

23<sup>rd</sup> European Symposium  
on Poultry Nutrition

ESPN  
2023

RMINI/TALY JUNE 21 - 24

## Feeding the microbiome

Richard Ducatelle, Evy Goossens, Venessa Eeckhout & Filip Van Immerseel  
Ghent University, Belgium

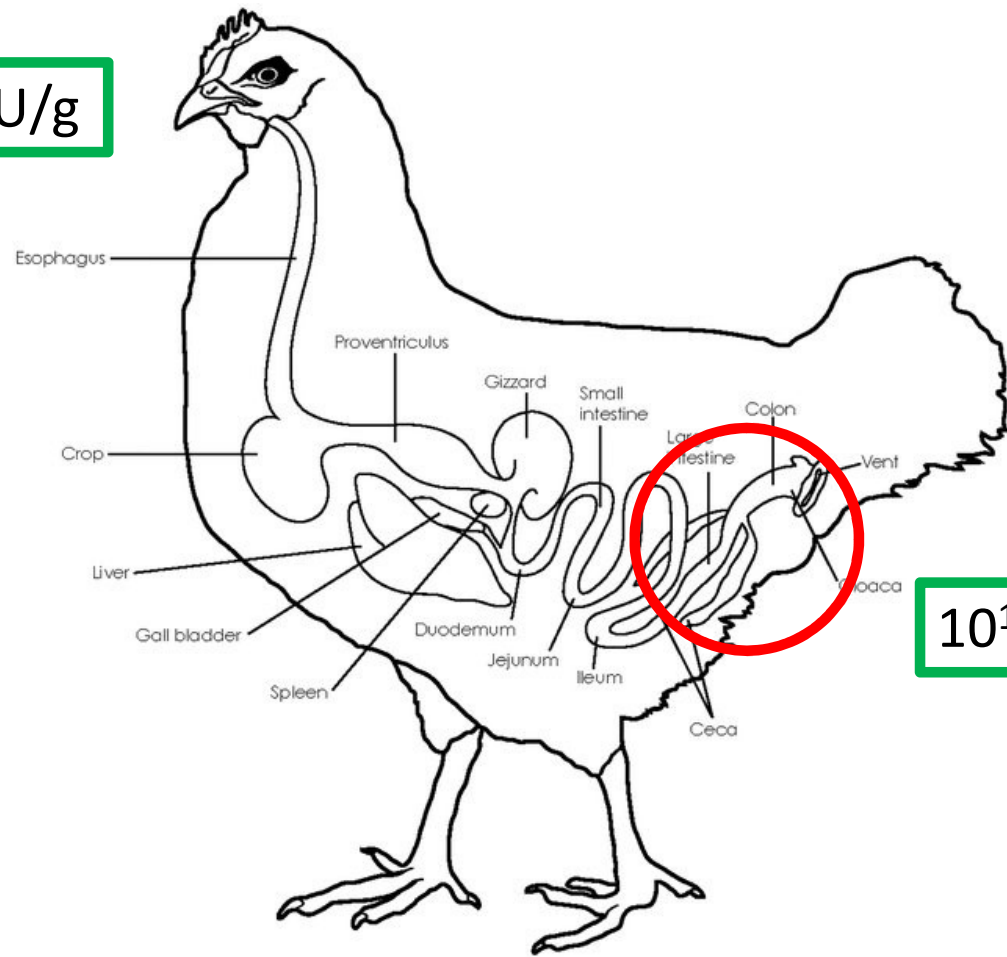




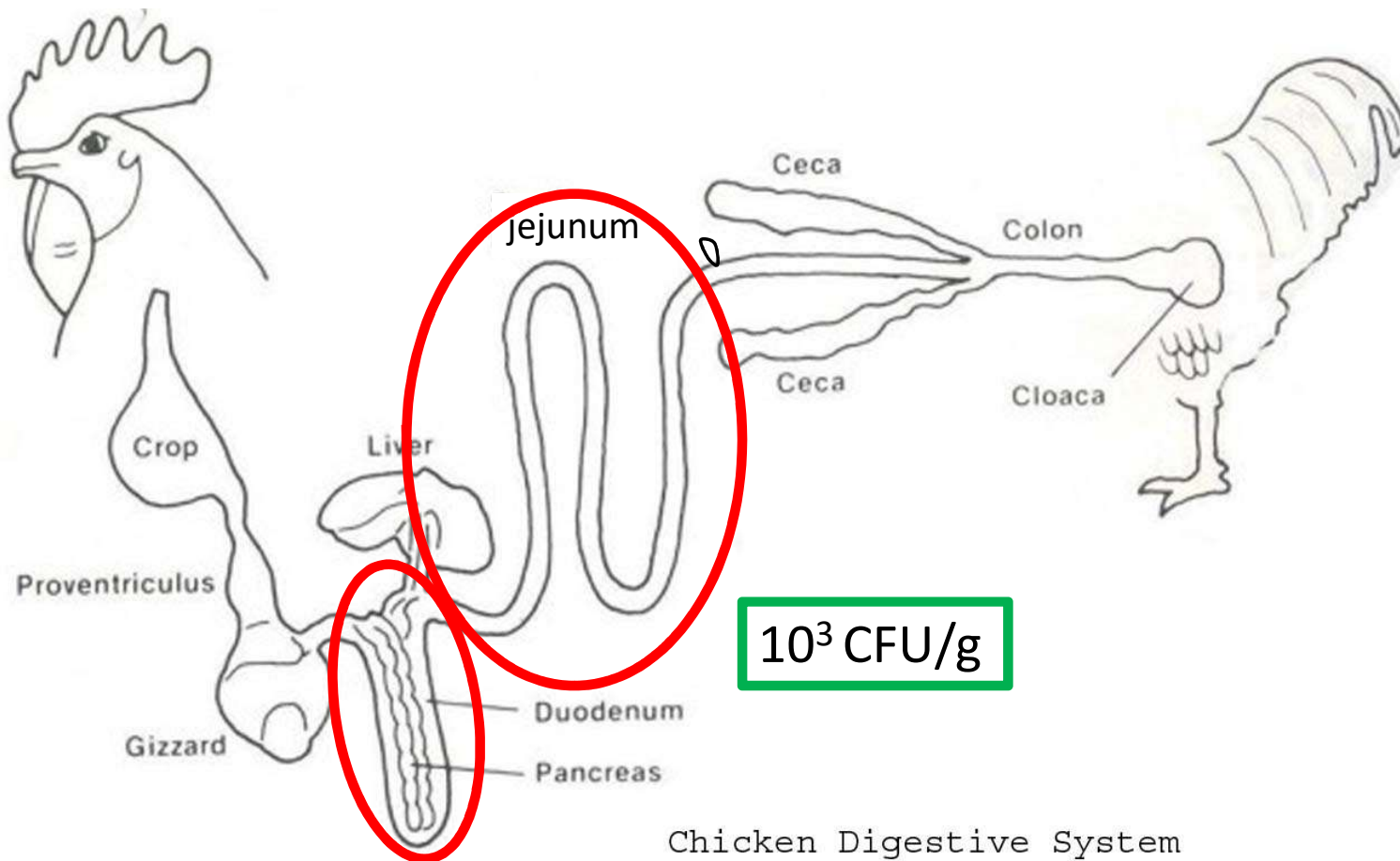
## VETERINARY SCIENCES



$10^6$  CFU/g

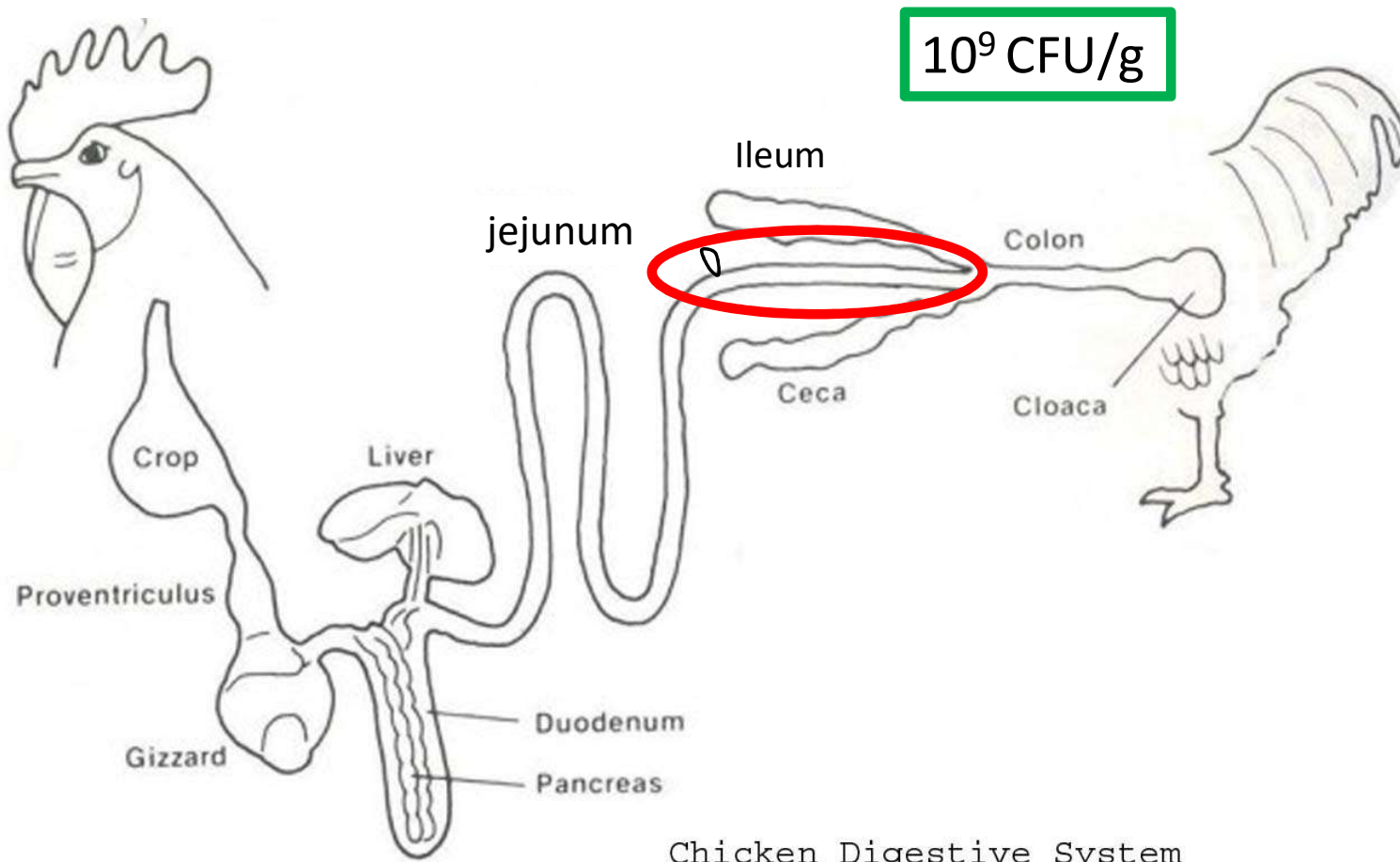


$10^{11}$  CFU/g

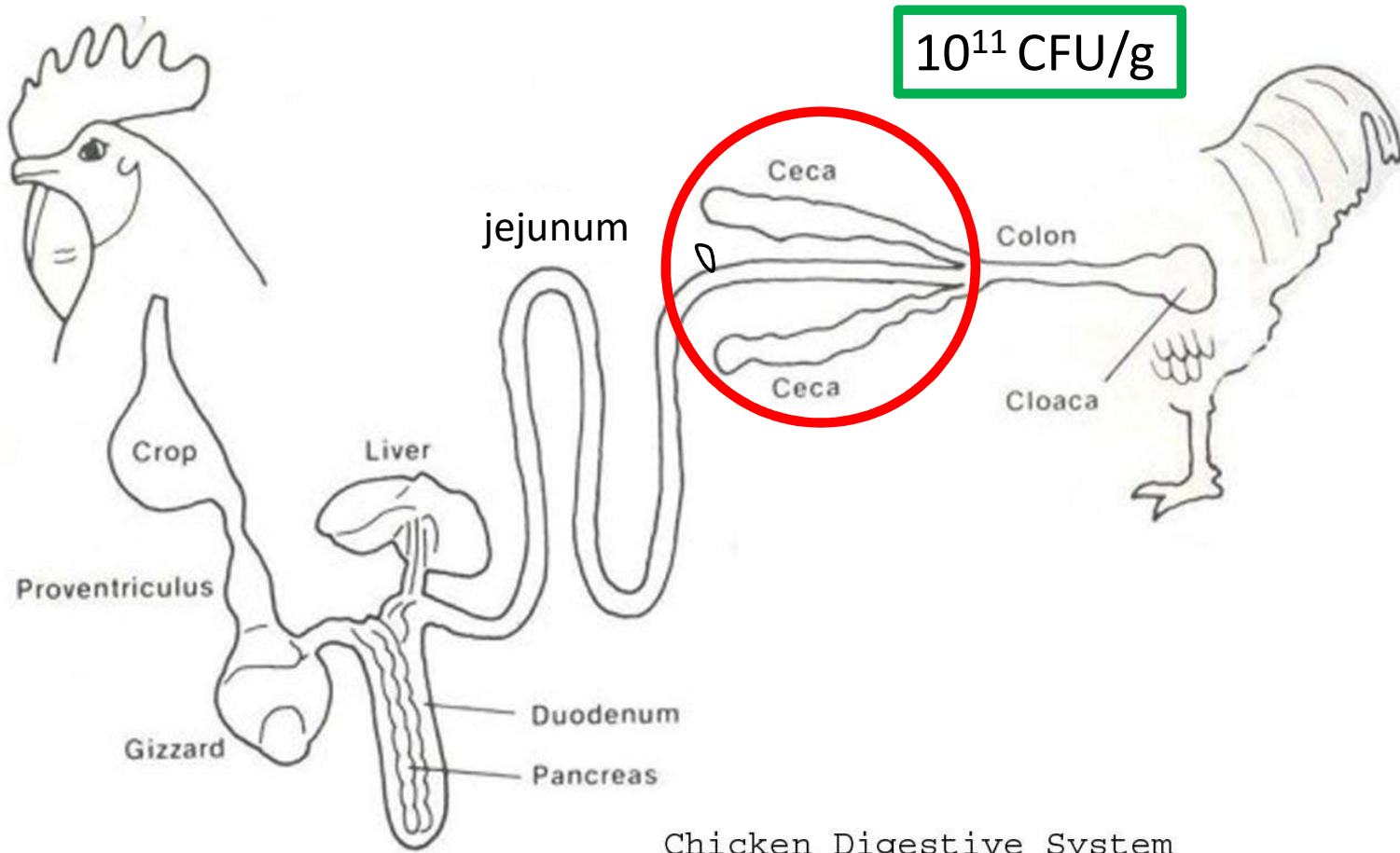


Chicken Digestive System





Chicken Digestive System



Chicken Digestive System

# The microbiota of the human colon

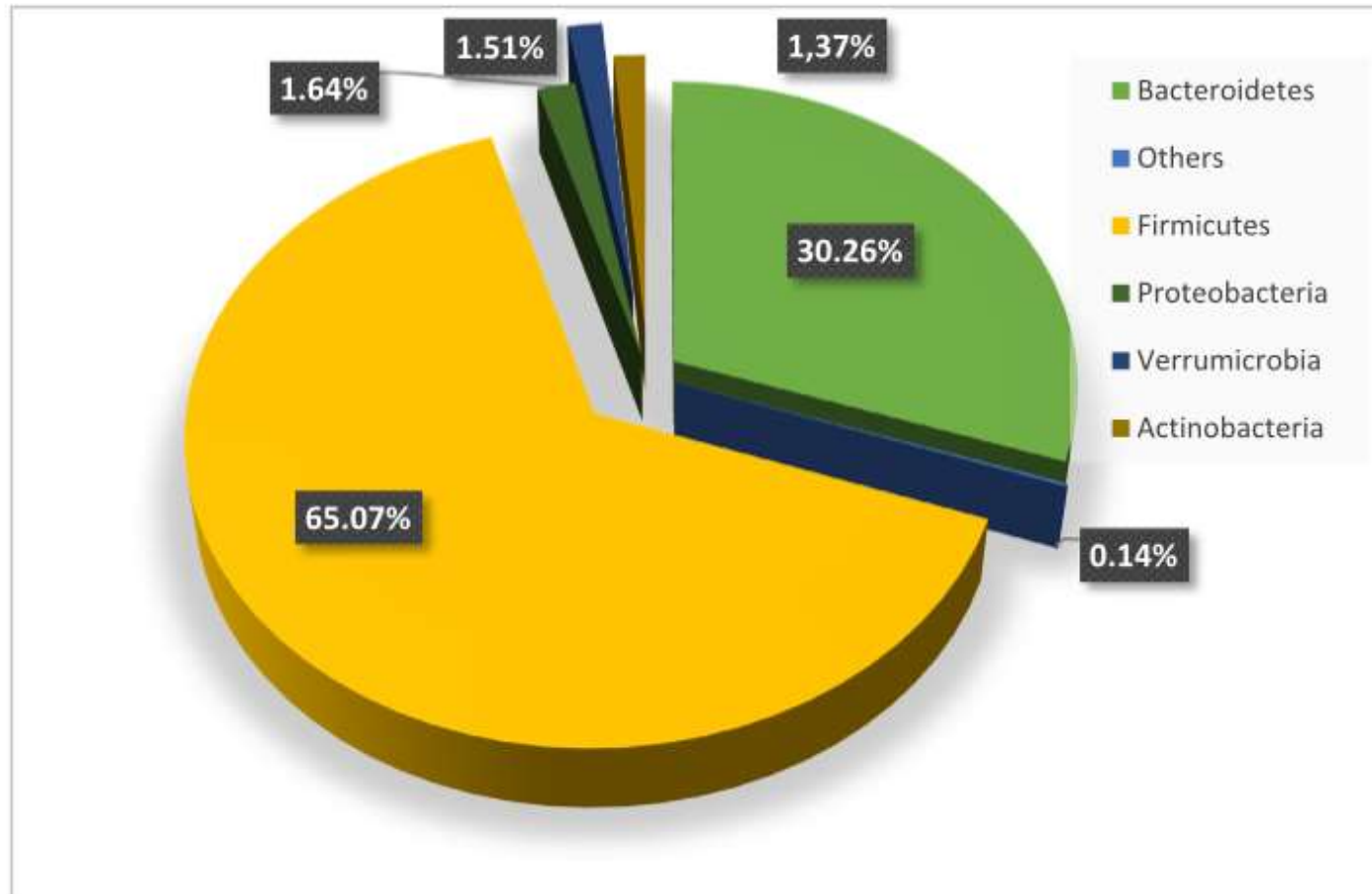
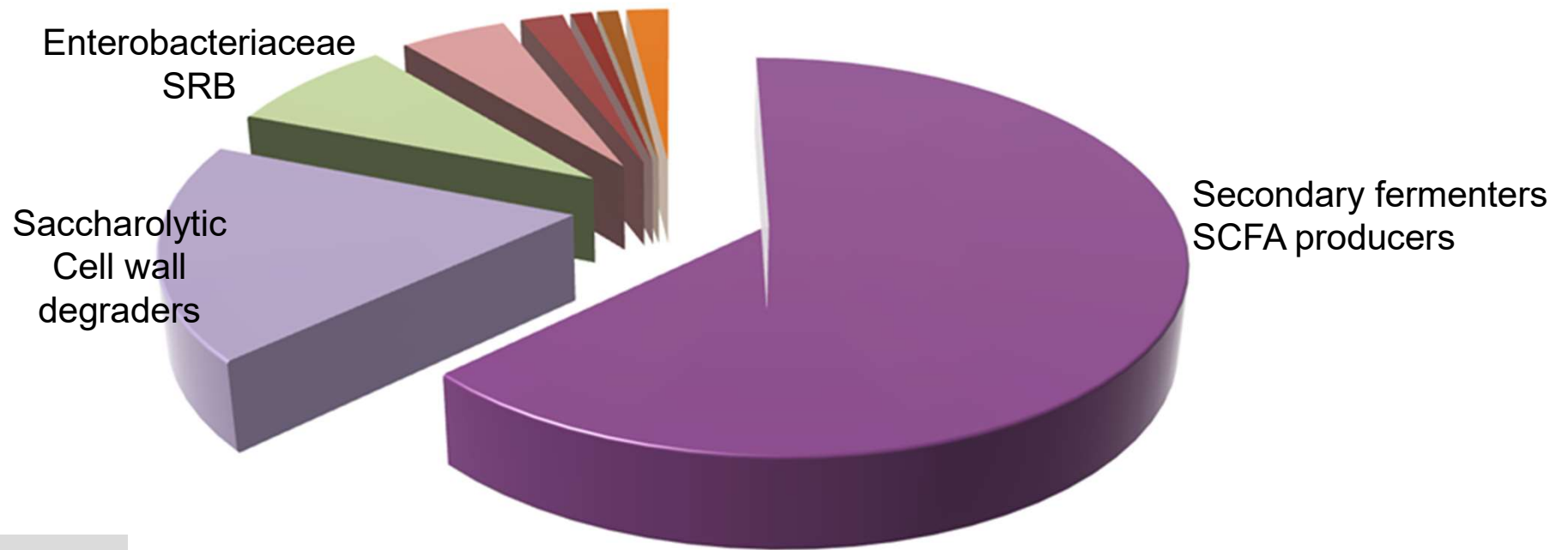


Fig. 2. Distribution of major bacterial phyla population according to their relative abundance in the human gut [28].

# The mature microbiota of the chicken caeca



Dominant phyla:

▣ *Firmicutes*

▣ *Bacteroidetes*

▣ *Proteobacteria*

▣ *Actinobacteria*

▣ *Verrucomicrobia*

▣ *Spirochaetes*

▣ *Fusobacteria*

▣ other



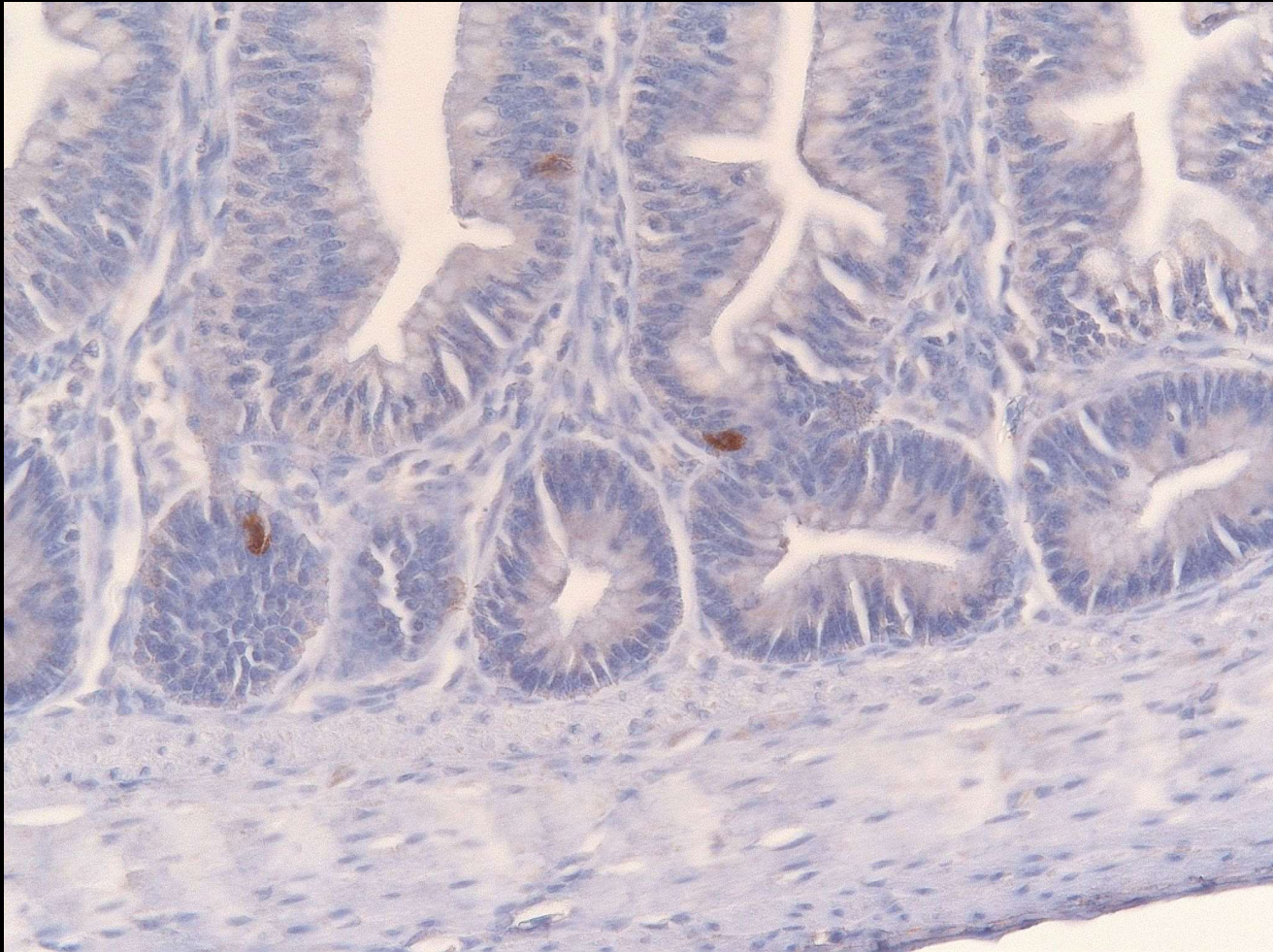
Short chain fatty acids, lactate, succinate, ....

**Contribution to total energy harvest = 10%**

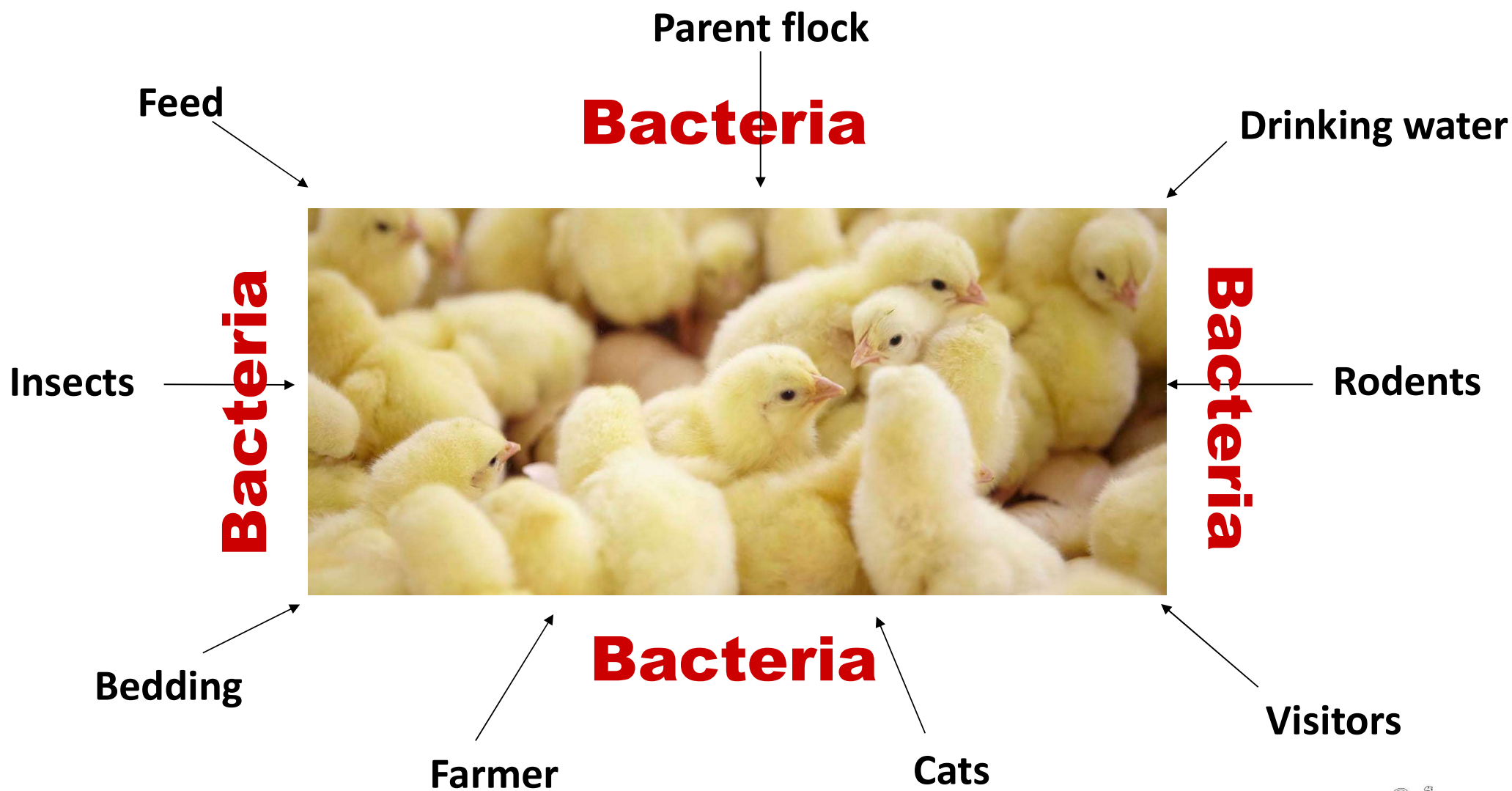
Effects of butyrate

Interkingdom signaling

L-cells produce GLP2



# Where does the microbiome come from?





# Who is feeding the microbiome?

The host !



Gastroenterology Report, 7(1), 2019, 3–12

doi: [10.1093/gastro/goy052](https://doi.org/10.1093/gastro/goy052)

Review

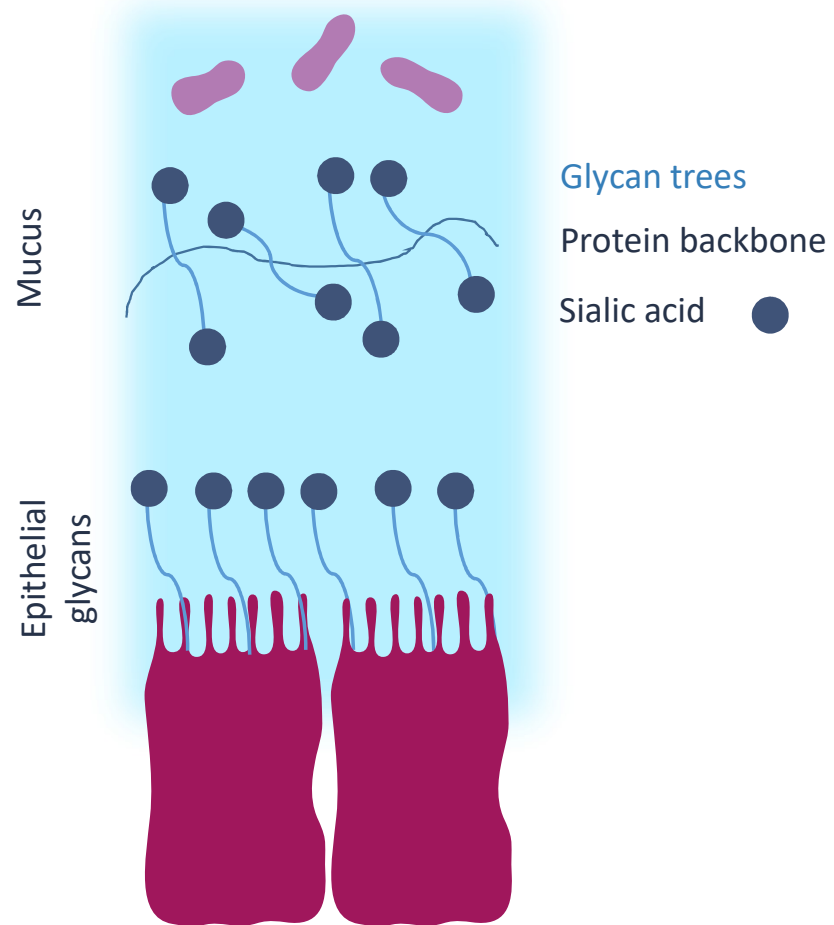
REVIEW

# Fight them or feed them: how the intestinal mucus layer manages the gut microbiota

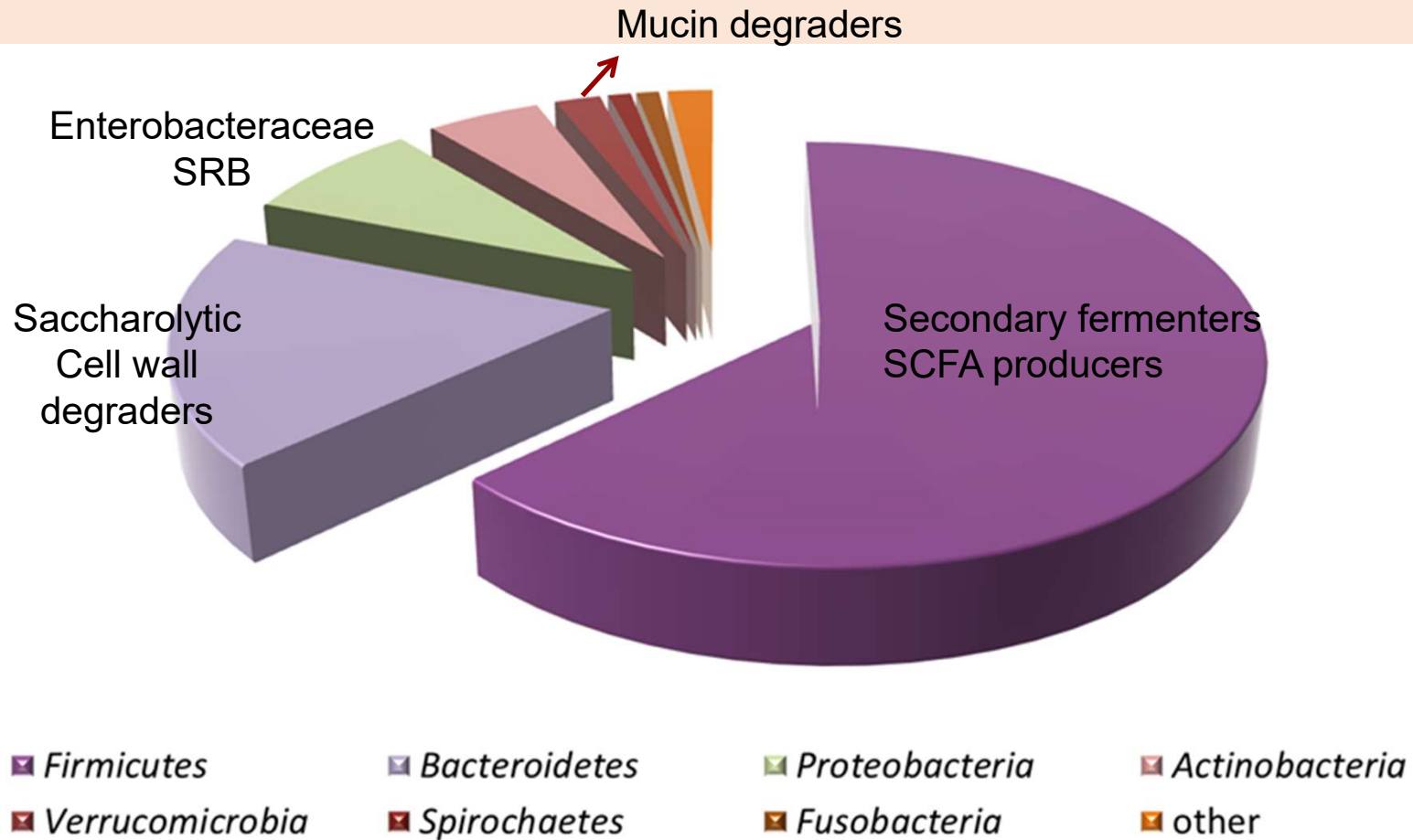
Bjoern O. Schroeder  \*

Wallenberg Laboratory and Sahlgrenska Center for Cardiovascular and Metabolic Research, Department of Molecular and Clinical Medicine, Institute of Medicine, Bruna Stråket 16, University of Gothenburg, SE 413 45 Gothenburg, Sweden

# Intestinal mucus



# The microbiota of the chicken gut





# Sialidases and fucosidases of *Akkermansia muciniphila* are crucial for growth on mucin and nutrient sharing with mucus-associated gut bacteria

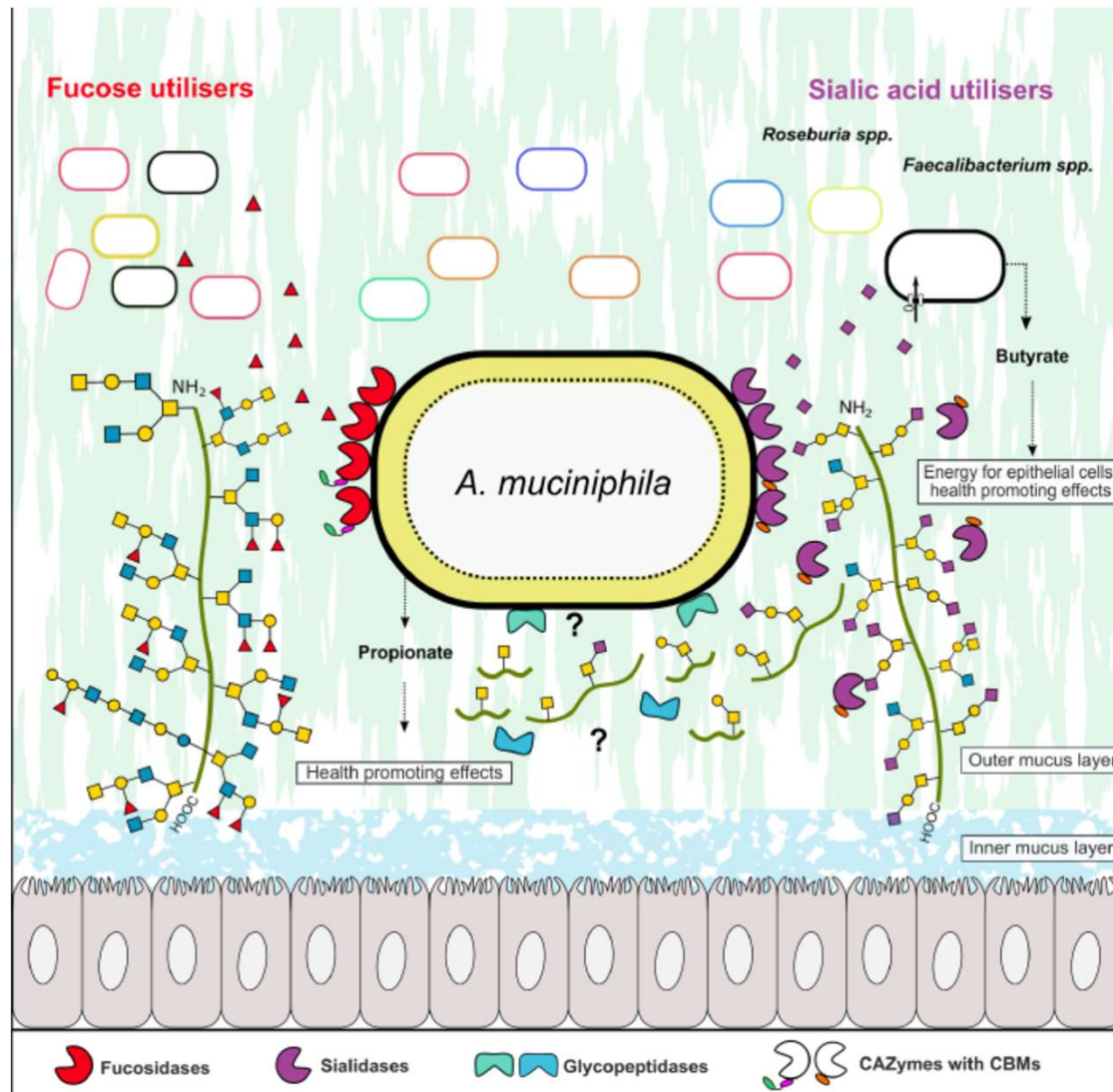
Received: 1 October 2022

Accepted: 21 March 2023

Published online: 01 April 2023

Bashar Shuoker<sup>1,2,7</sup>, Michael J. Pichler<sup>1,7</sup>, Chunsheng Jin<sup>3</sup>, Hiroka Sakanaka<sup>1</sup>, Haiyang Wu<sup>4</sup>, Ana Martínez Gascueña<sup>4</sup>, Jining Liu<sup>5</sup>, Tine Sofie Nielsen<sup>1</sup>, Jan Holgersson<sup>5</sup>, Eva Nordberg Karlsson<sup>2</sup>, Nathalie Juge<sup>4</sup>, Sebastian Meier<sup>6</sup>, Jens Preben Morth<sup>1</sup>✉, Niclas G. Karlsson<sup>3</sup> & Maher Abou Hachem<sup>1</sup>✉





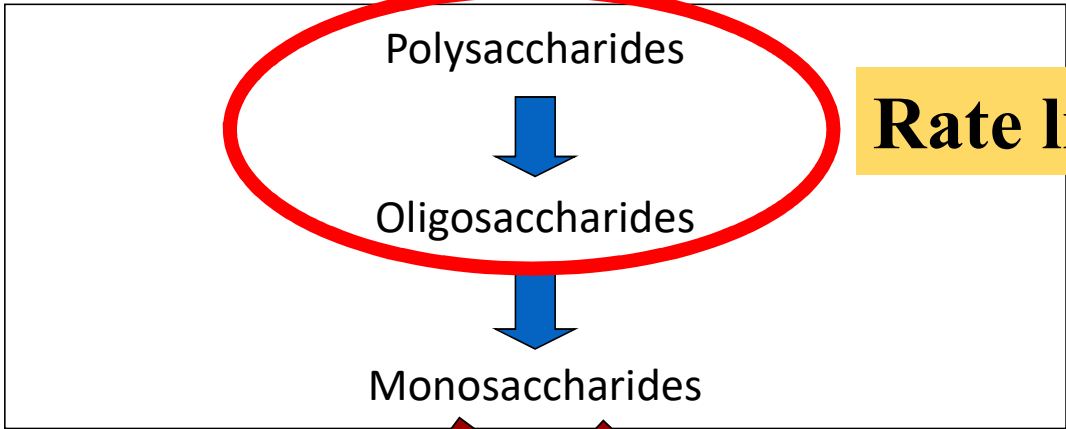
(Shuoker et al., 2023)

# Feeding the microbiome?

The leftovers !

**Dietary fiber = NSP**

**Residual protein**



**Rate limiting steps !**

Bacteroidetes, lactobacilli, bifidobacteria, ...

Lactate, acetate, succinate

H<sub>2</sub>

Firmicutes

*Clostridium* cluster IX

Propionate

Firmicutes  
*Clostridium* cluster IV and XIVa

Butyrate

Methanogenic bacteria

CH<sub>4</sub> (Archaeobacteria)

# Dietary fiber

The portion of plant-derived food that cannot be completely broken down by [digestive enzymes](#)

Planta (2016) 243:281–296  
DOI 10.1007/s00425-015-2450-x



REVIEW

## The polyploidy and its key role in plant breeding

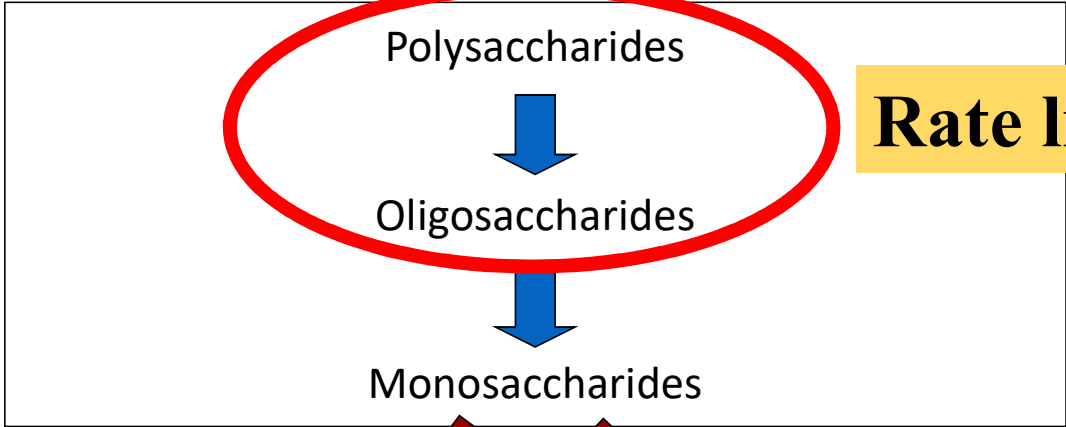
Mariana Cansian Sattler<sup>1</sup> · Carlos Roberto Carvalho<sup>2</sup> · Wellington Ronildo Clarindo<sup>1</sup>

**Higher yields**

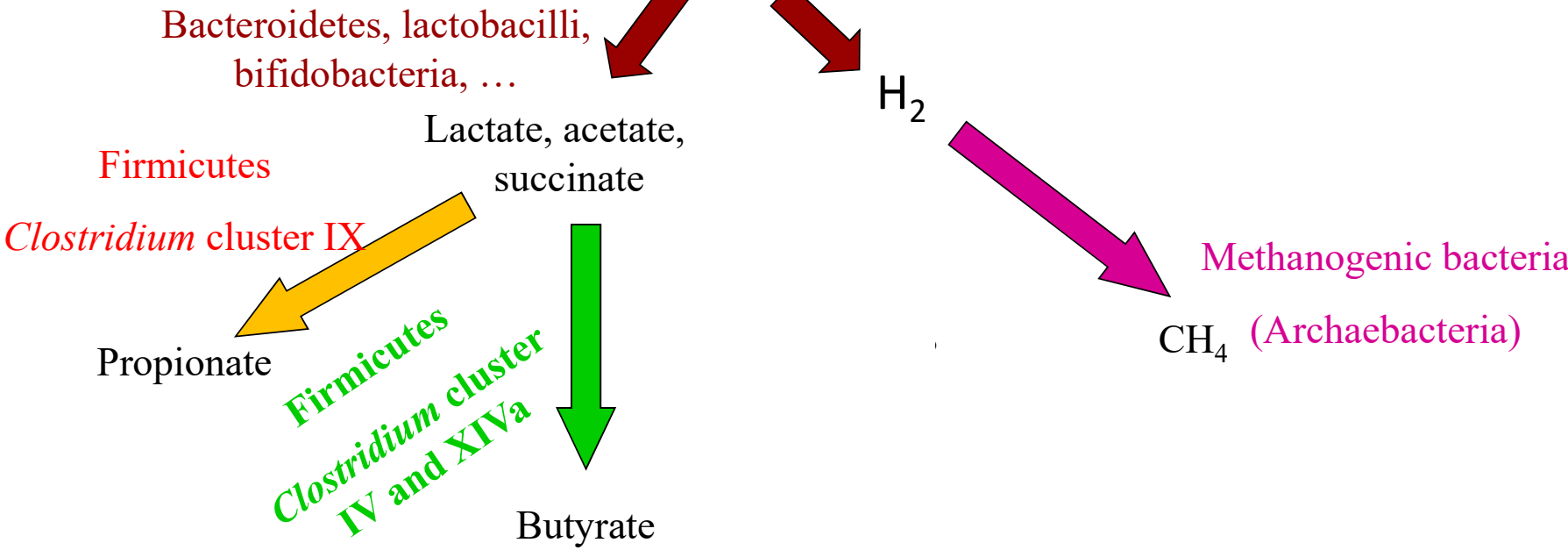
**Increased cell size**

**Less dietary fiber**

**Dietary fiber = NSP**

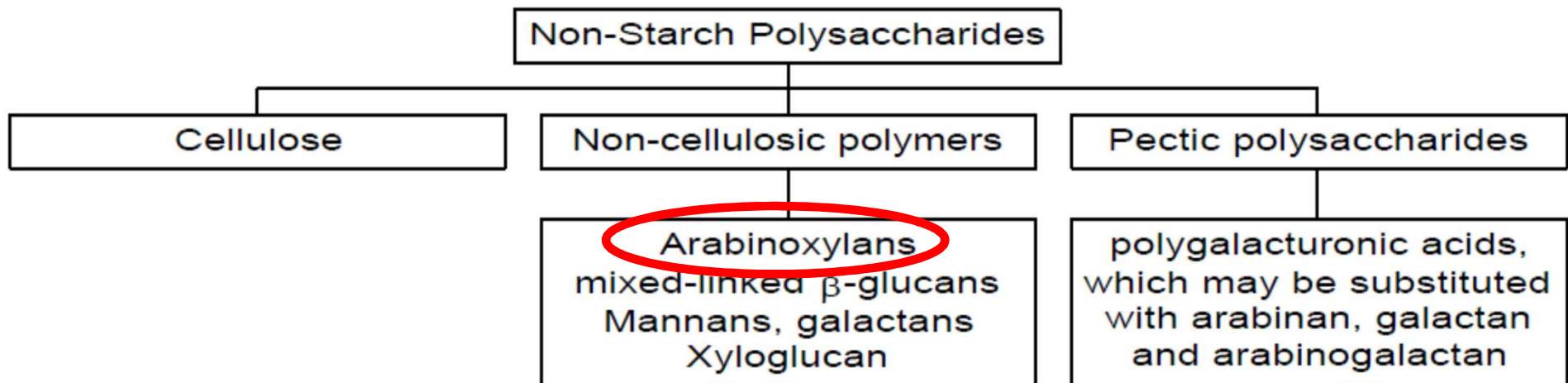


**Rate limiting steps !**





# Dietary fiber =

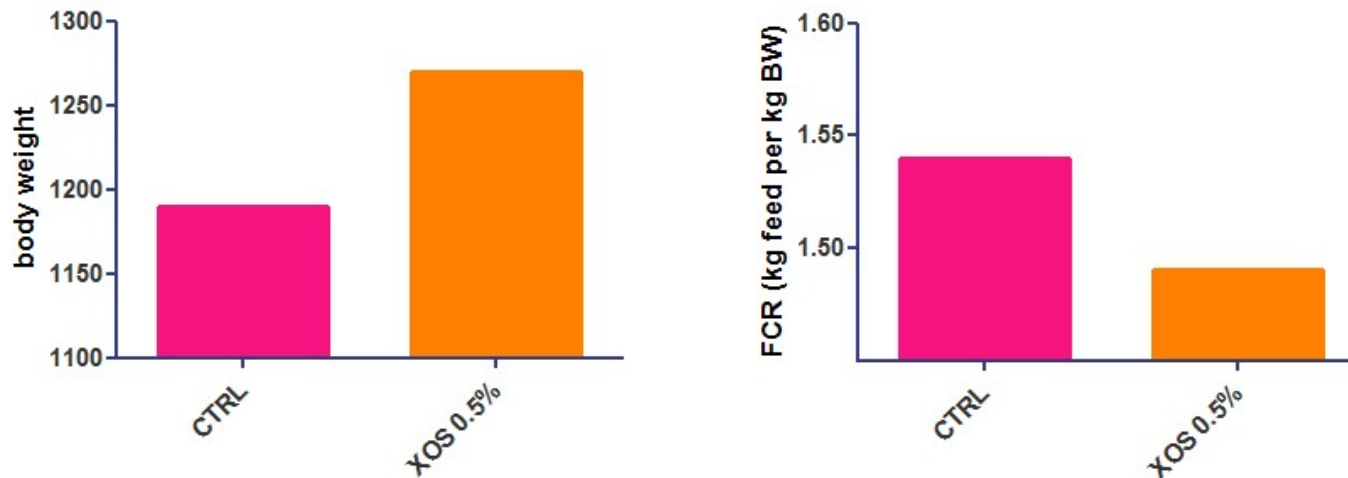


(Choct M., 1997)

# Effects of oligosaccharide supplements: the case of XOS

Add 0.5% XOS to broiler feed

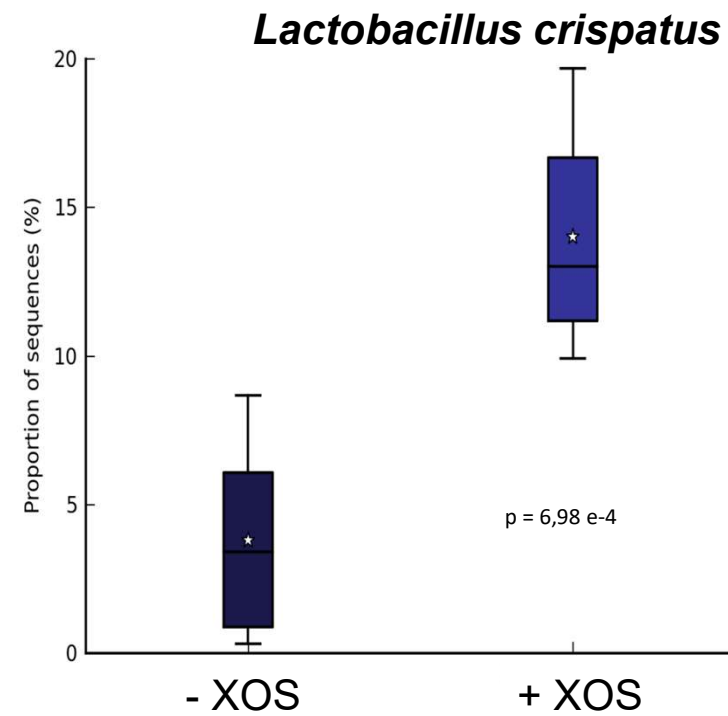
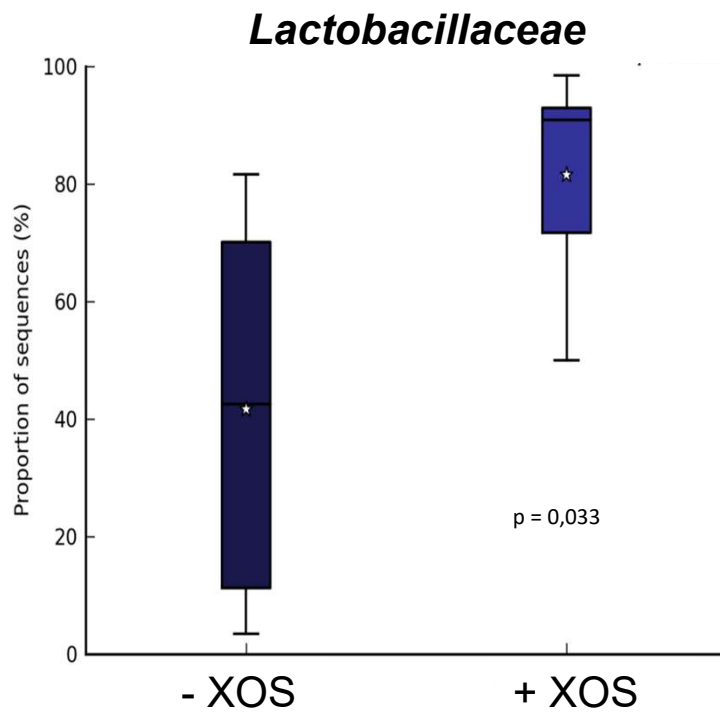
**Day 26:**



De Maesschalck et al., 2015

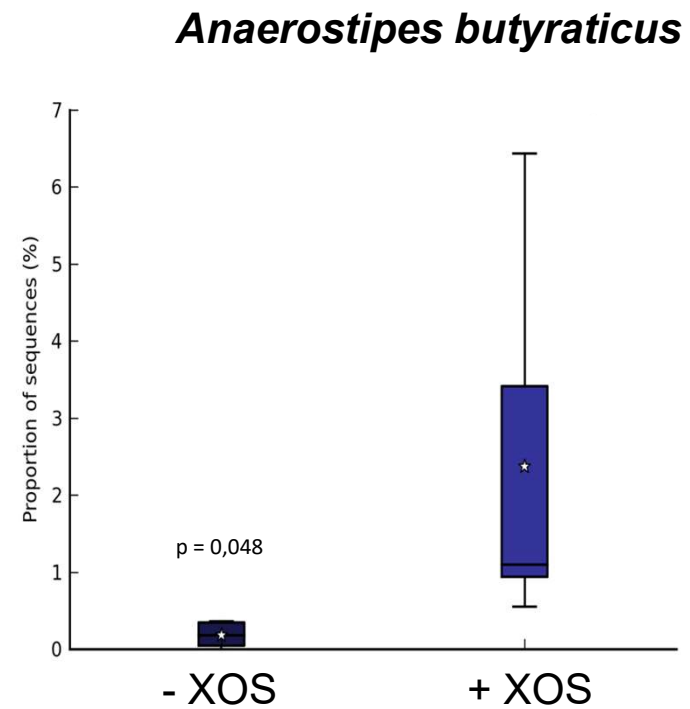
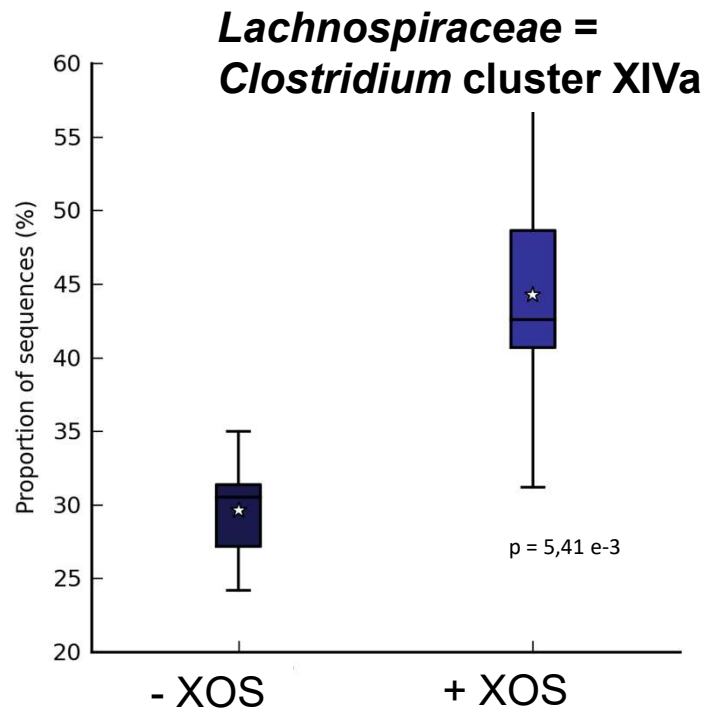
# Microbiota

## Colon

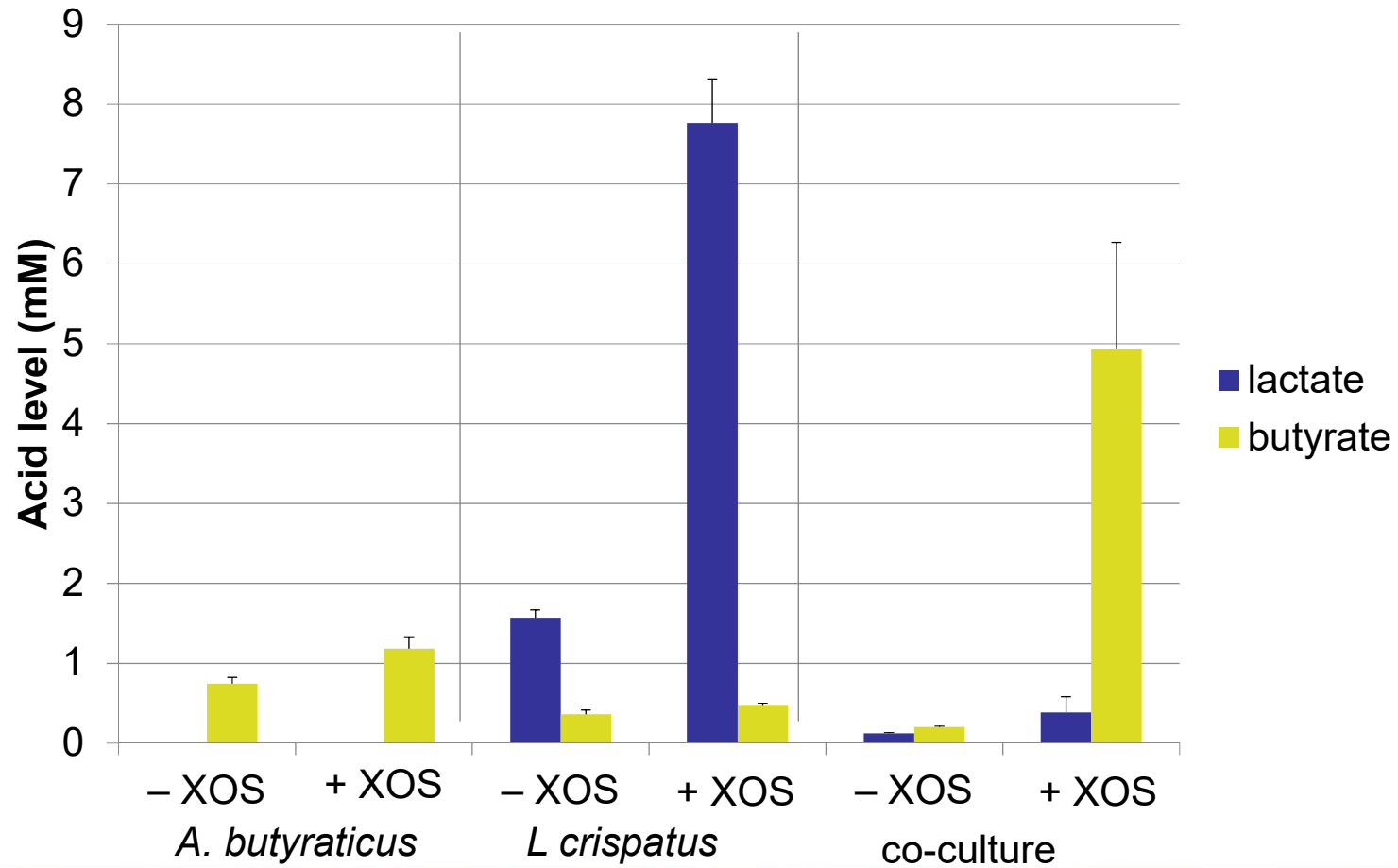


# Microbiota

## Caecum



# Cross-feeding

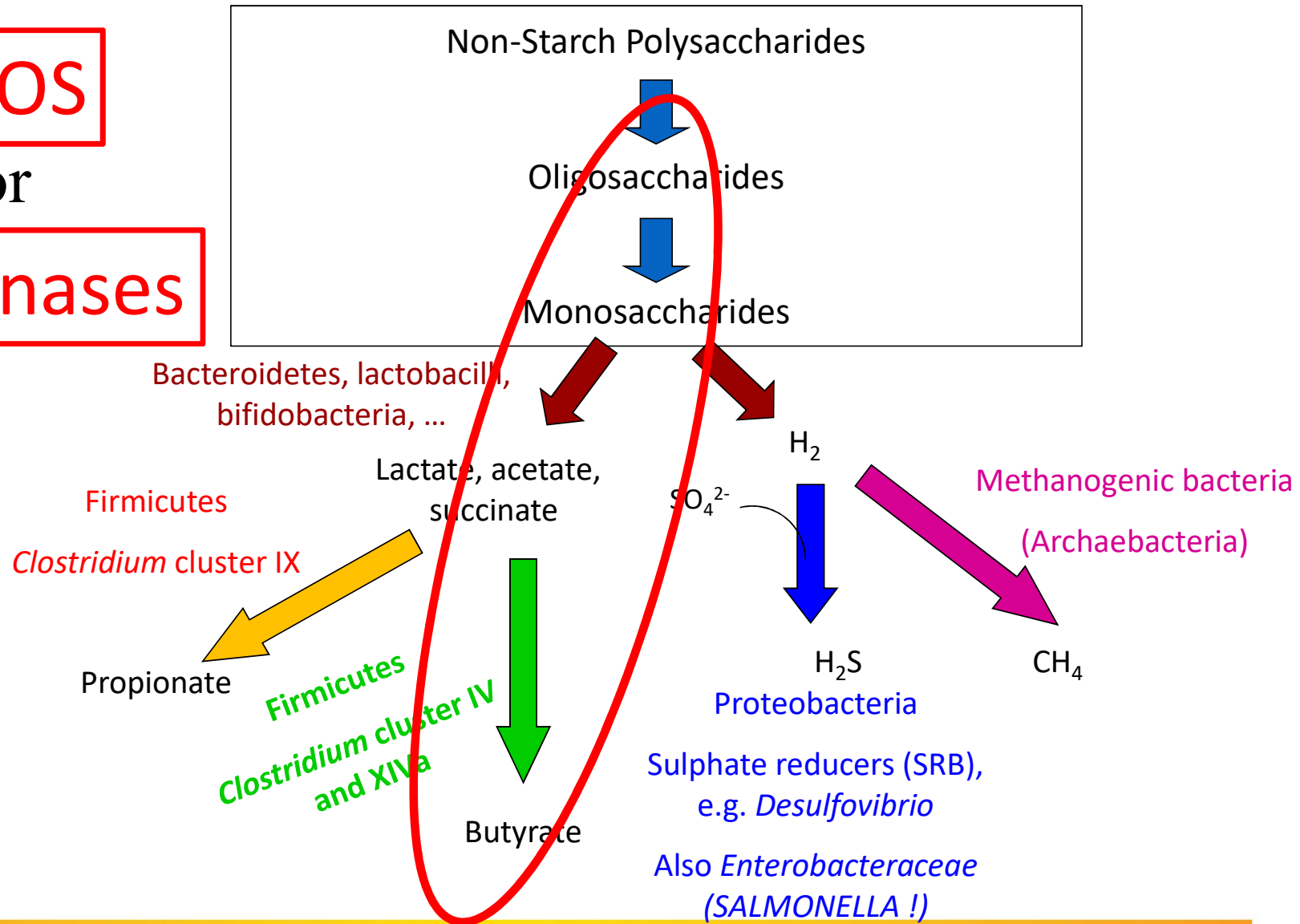




+ XOS

or

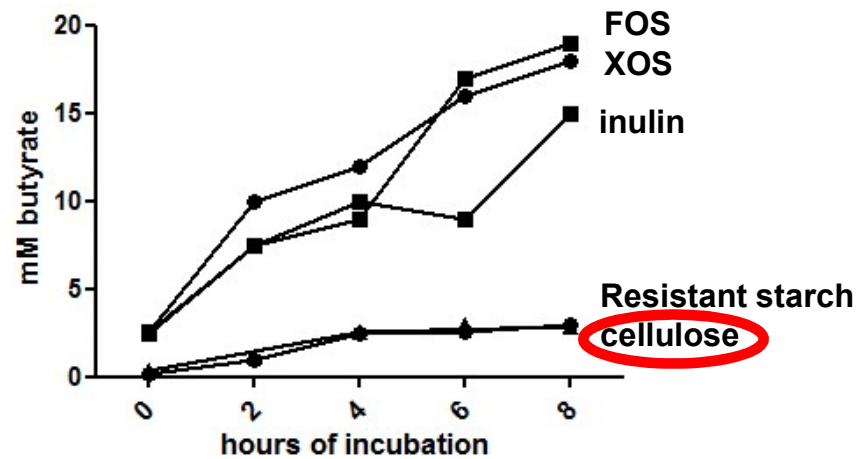
+ xylanases



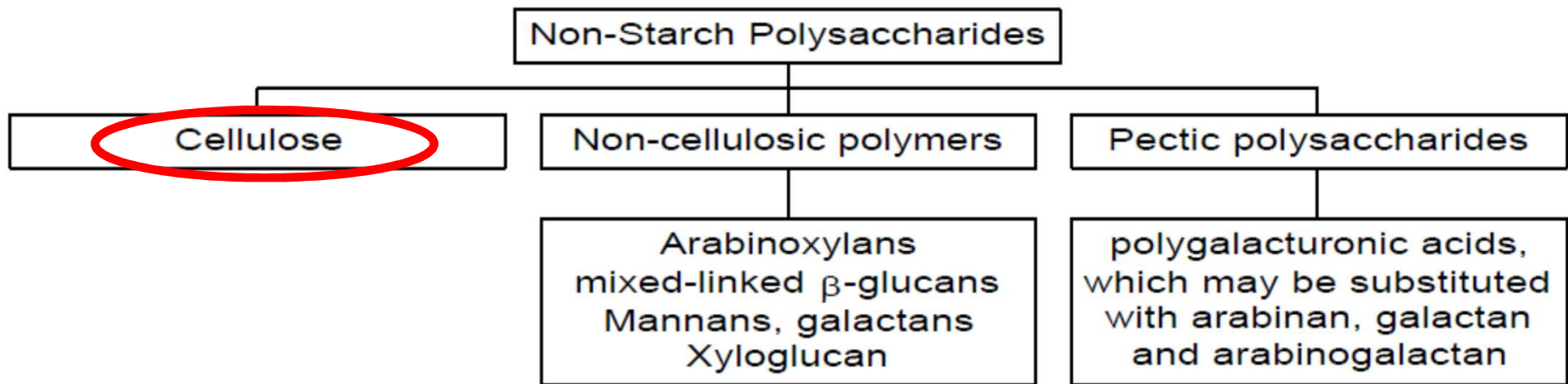
# Effects of different prebiotics on cecal butyrate producing microbiota



+ substrates  
Measure SCFA during time

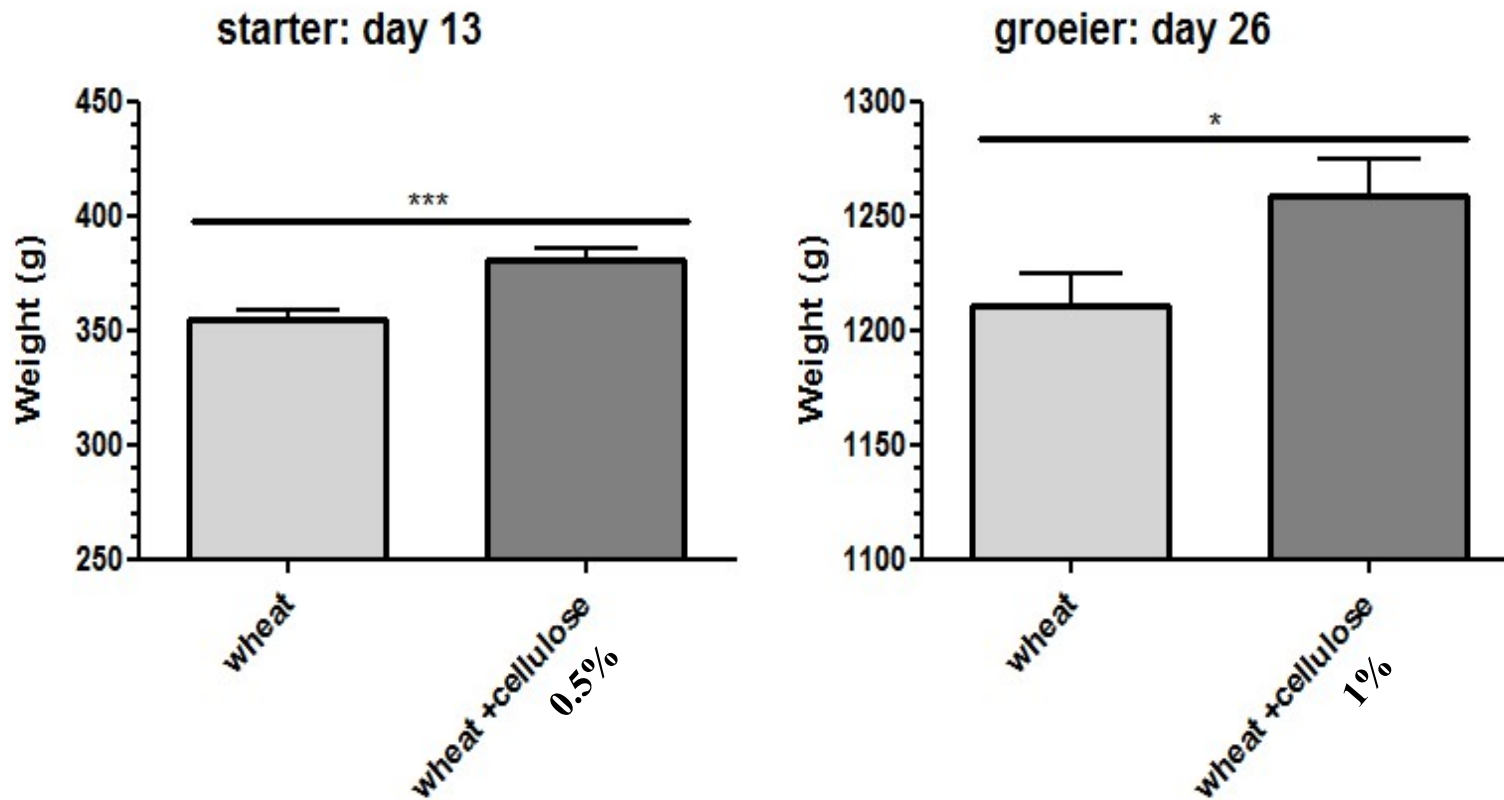


# Dietary fiber =



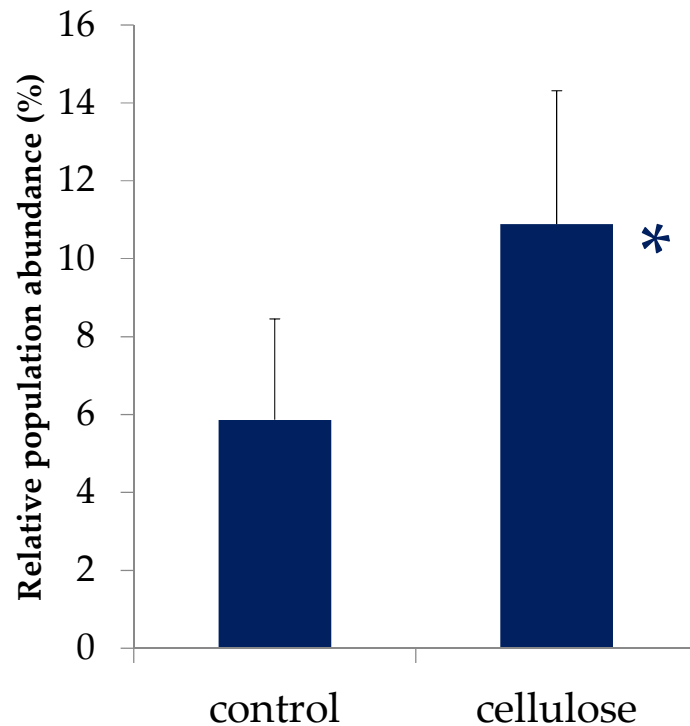
(Choct M., 1997)

# *in vivo* effects of cellulose



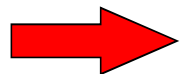
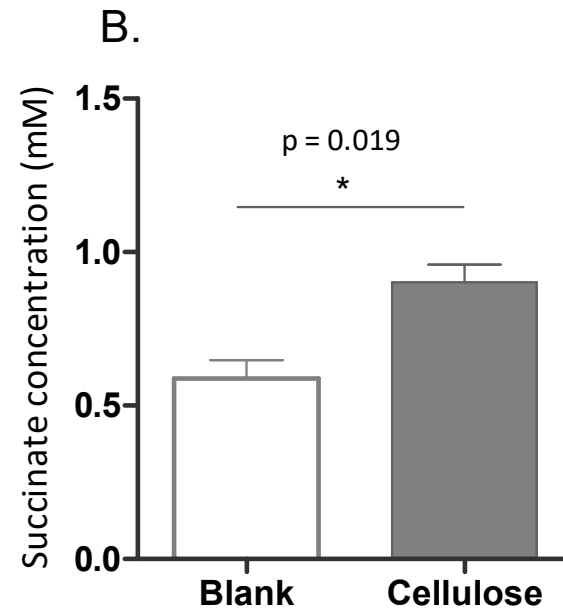
# Microbiota composition: caeca

## *Alistipes*

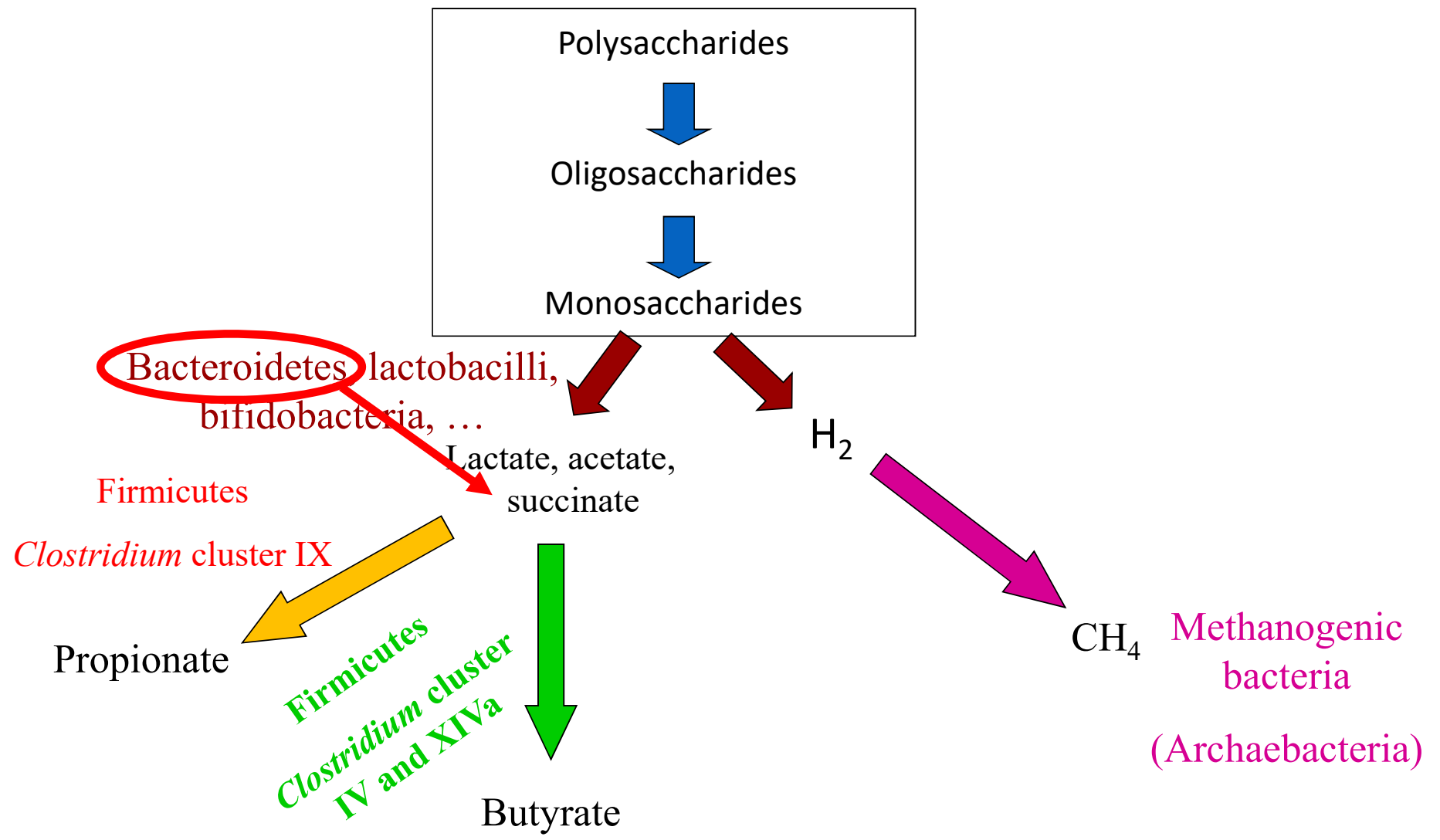


phylum: *Bacteroidetes*  
class: *Bacteroidia*  
order: *Bacteroidales*  
family: *Rikenellaceae*  
genus: *Alistipes*

# Succinate production by *Alistipes finegoldii* in the presence of cellulose

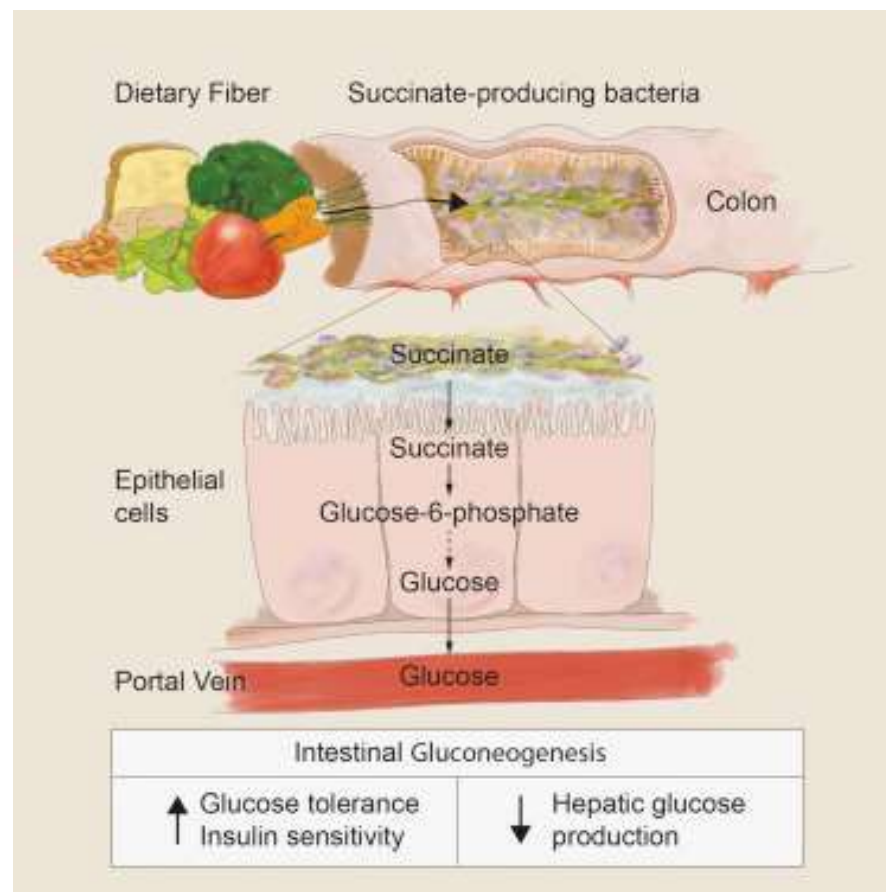


Cellulose breakdown in the intestinal lumen does not require a microbial network





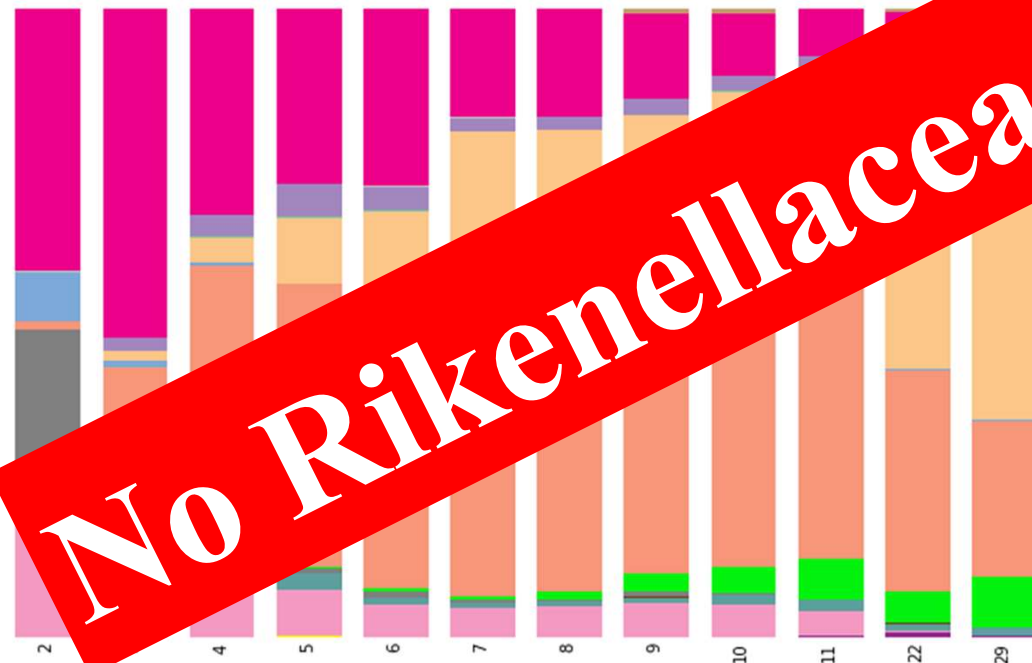
## Microbiota-Produced Succinate Improves Glucose Homeostasis via Intestinal Gluconeogenesis



Filipe De Vadder, Petia Kovatcheva-Datchary, Carine Zitoun, Adeline Duchamp, Fredrik Bäckhed, Gilles Mithieux

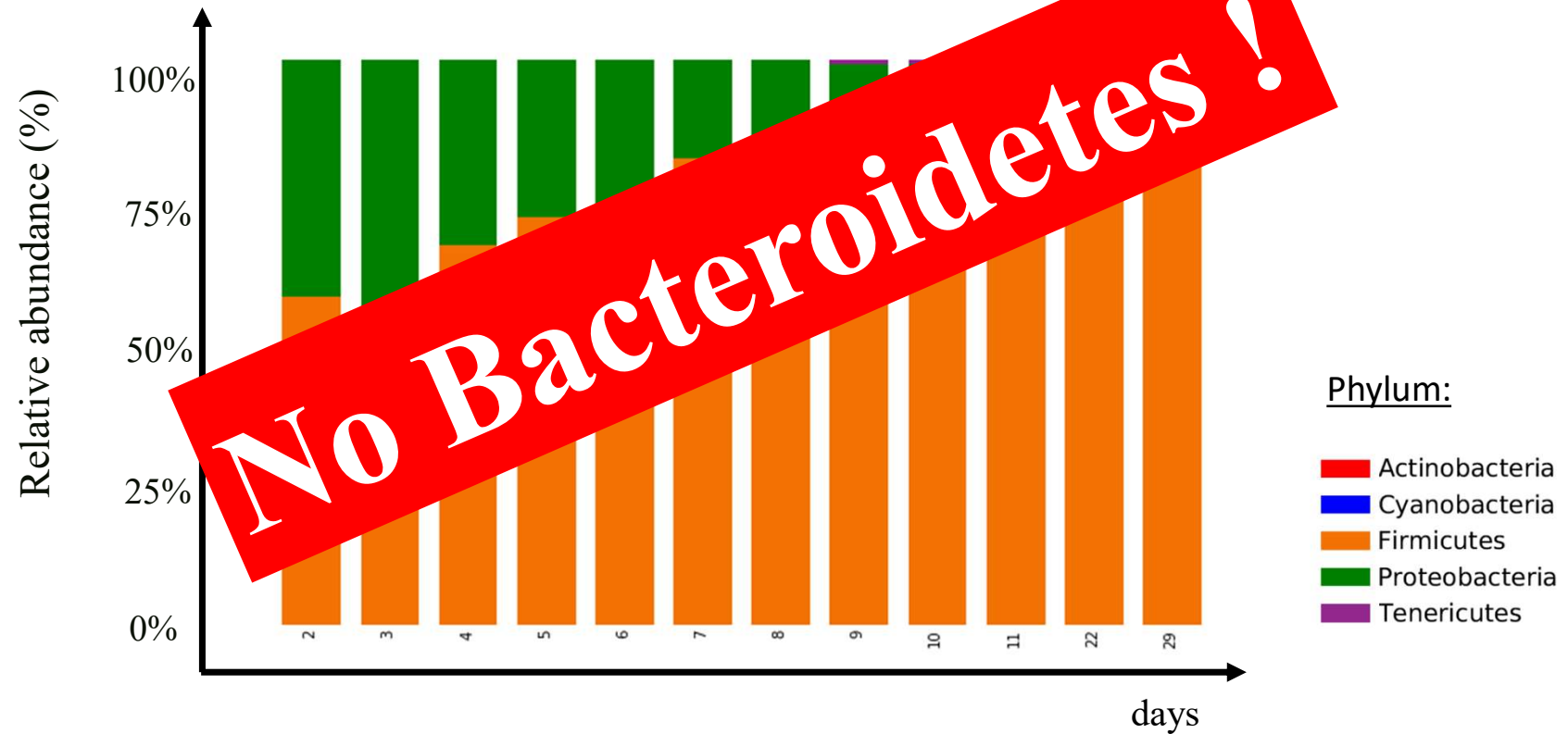
Cell Metabolism, Volume 24, Issue 1, 2016, 151–157

# Relative abundance of the families

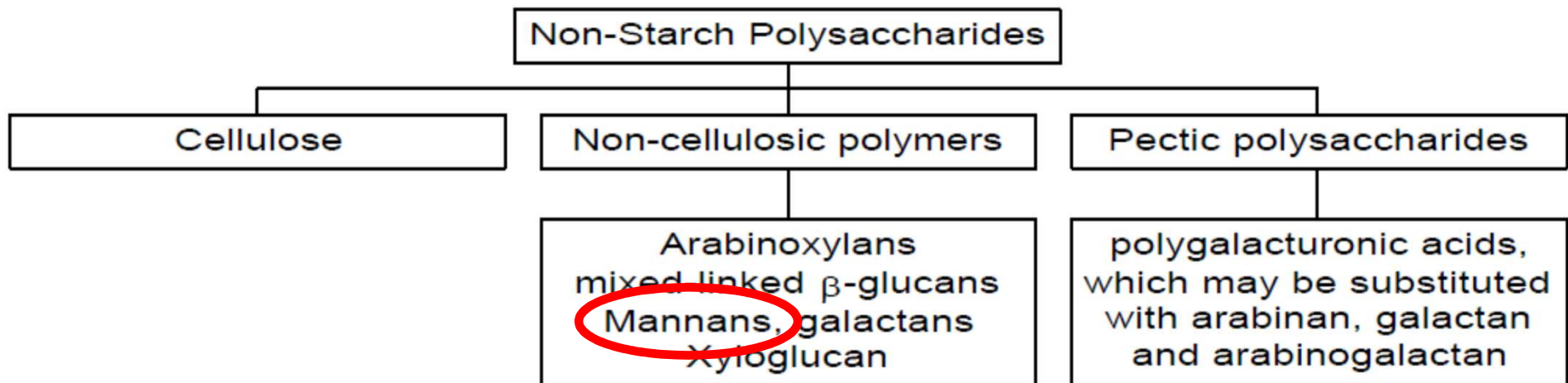


- D\_0\_Bacteria;D\_1\_Actinobacteria;D\_2\_Actinobacteria;D\_3\_Corynebacteriales;D\_4\_Corynebacteriaceae
- D\_0\_Bacteria;D\_1\_Actinobacteria;D\_2\_Actinobacteria;D\_3\_Micrococcales;D\_4\_Micrococcaceae
- D\_0\_Bacteria;D\_1\_Actinobacteria;D\_2\_Coriobacterii;D\_3\_Coriobacteriales;D\_4\_Eggerthellaceae
- D\_0\_Bacteria;D\_1\_Cyanobacteria;D\_2\_Oxyphotobacteria;D\_3\_Chloroplast;D\_4\_Phaseolus acutifolius (teary bean)
- D\_0\_Bacteria;D\_1\_Firmicutes;D\_2\_Bacilli;D\_3\_Bacillales;D\_4\_Bacillaceae
- D\_0\_Bacteria;D\_1\_Firmicutes;D\_2\_Bacilli;D\_3\_Bacillales;D\_4\_Paenibacillaceae
- D\_0\_Bacteria;D\_1\_Firmicutes;D\_2\_Bacilli;D\_3\_Bacillales;D\_4\_Staphylococcaceae
- D\_0\_Bacteria;D\_1\_Firmicutes;D\_2\_Bacilli;D\_3\_Lactobacillales;D\_4\_Enterococcaceae
- D\_0\_Bacteria;D\_1\_Firmicutes;D\_2\_Bacilli;D\_3\_Lactobacillales;D\_4\_Lactobacillaceae
- D\_0\_Bacteria;D\_1\_Firmicutes;D\_2\_Clostridia;D\_3\_Clostridiales;D\_4\_Christensenellaceae
- D\_0\_Bacteria;D\_1\_Firmicutes;D\_2\_Clostridia;D\_3\_Clostridiales;D\_4\_Clostridiaceae 1
- D\_0\_Bacteria;D\_1\_Firmicutes;D\_2\_Clostridia;D\_3\_Clostridiales;D\_4\_Clostridiales vadinBB60 group
- D\_0\_Bacteria;D\_1\_Firmicutes;D\_2\_Clostridia;D\_3\_Clostridiales;D\_4\_Lachnospiraceae
- D\_0\_Bacteria;D\_1\_Firmicutes;D\_2\_Clostridia;D\_3\_Clostridiales;D\_4\_Peptostreptococcaceae
- D\_0\_Bacteria;D\_1\_Firmicutes;D\_2\_Clostridia;D\_3\_Clostridiales;D\_4\_Ruminococcaceae
- D\_0\_Bacteria;D\_1\_Firmicutes;D\_2\_Clostridia;D\_3\_Clostridiales;Other
- D\_0\_Bacteria;D\_1\_Firmicutes;D\_2\_Erysipelotrichia;D\_3\_Erysipelotrichales;D\_4\_Erysipelotrichaceae
- D\_0\_Bacteria;D\_1\_Proteobacteria;D\_2\_Alphaproteobacteria;D\_3\_Rickettsiales;D\_4\_Mitochondria
- D\_0\_Bacteria;D\_1\_Proteobacteria;D\_2\_Gammaproteobacteria;D\_3\_Betaproteobacteriales;D\_4\_Burkholderiaceae
- D\_0\_Bacteria;D\_1\_Proteobacteria;D\_2\_Gammaproteobacteria;D\_3\_Enterobacteriales;D\_4\_Enterobacteriaceae
- D\_0\_Bacteria;D\_1\_Proteobacteria;D\_2\_Gammaproteobacteria;D\_3\_Pseudomonadales;D\_4\_Pseudomonadaceae
- D\_0\_Bacteria;D\_1\_Tenericutes;D\_2\_Mollicutes;D\_3\_Mollicutes RF39;D\_4\_uncultured bacterium

## Relative abundance of the phyla



# Dietary fiber =



(Choct M., 1997)



## Human Gut *Faecalibacterium prausnitzii* Deploys a Highly Efficient Conserved System To Cross-Feed on $\beta$ -Mannan-Derived Oligosaccharides

Lars J. Lindstad,<sup>a</sup> Galiana Lo,<sup>b</sup> Shaun Leivers,<sup>a</sup> Zijia Lu,<sup>c</sup> Leszek Michalak,<sup>a</sup> Gabriel V. Pereira,<sup>d</sup> Åsmund K. Røhr,<sup>a</sup> Eric C. Martens,<sup>d</sup> Lauren S. McKee,<sup>c</sup> Petra Louis,<sup>b</sup> Sylvia H. Duncan,<sup>b</sup> Bjørge Westereng,<sup>a</sup> Phillip B. Pope,<sup>a,e</sup> Sabina Leanti La Rosa<sup>a,e</sup>

### Primary degraders:

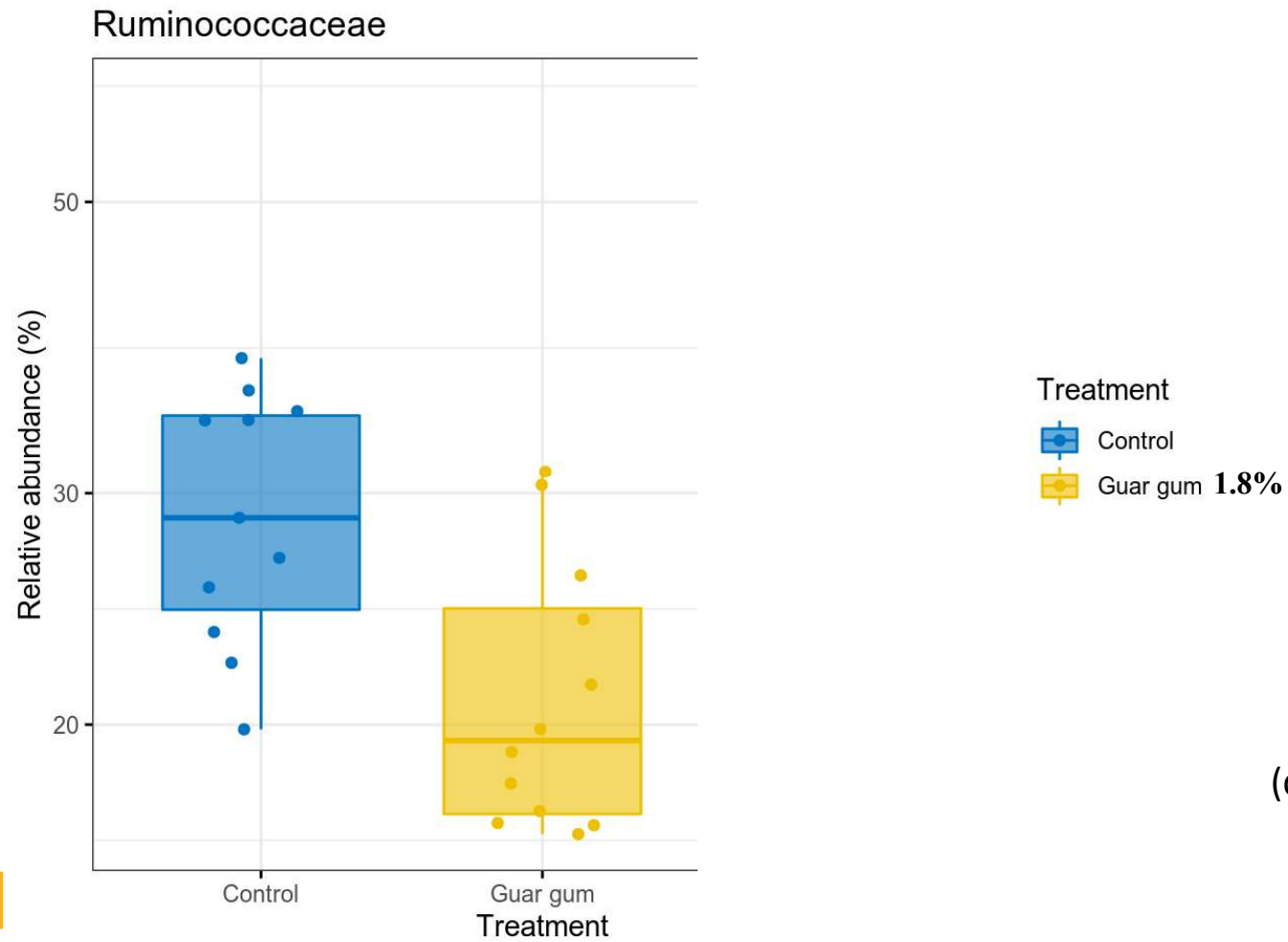
*Bacteroides ovatus*

*Roseburia intestinalis*



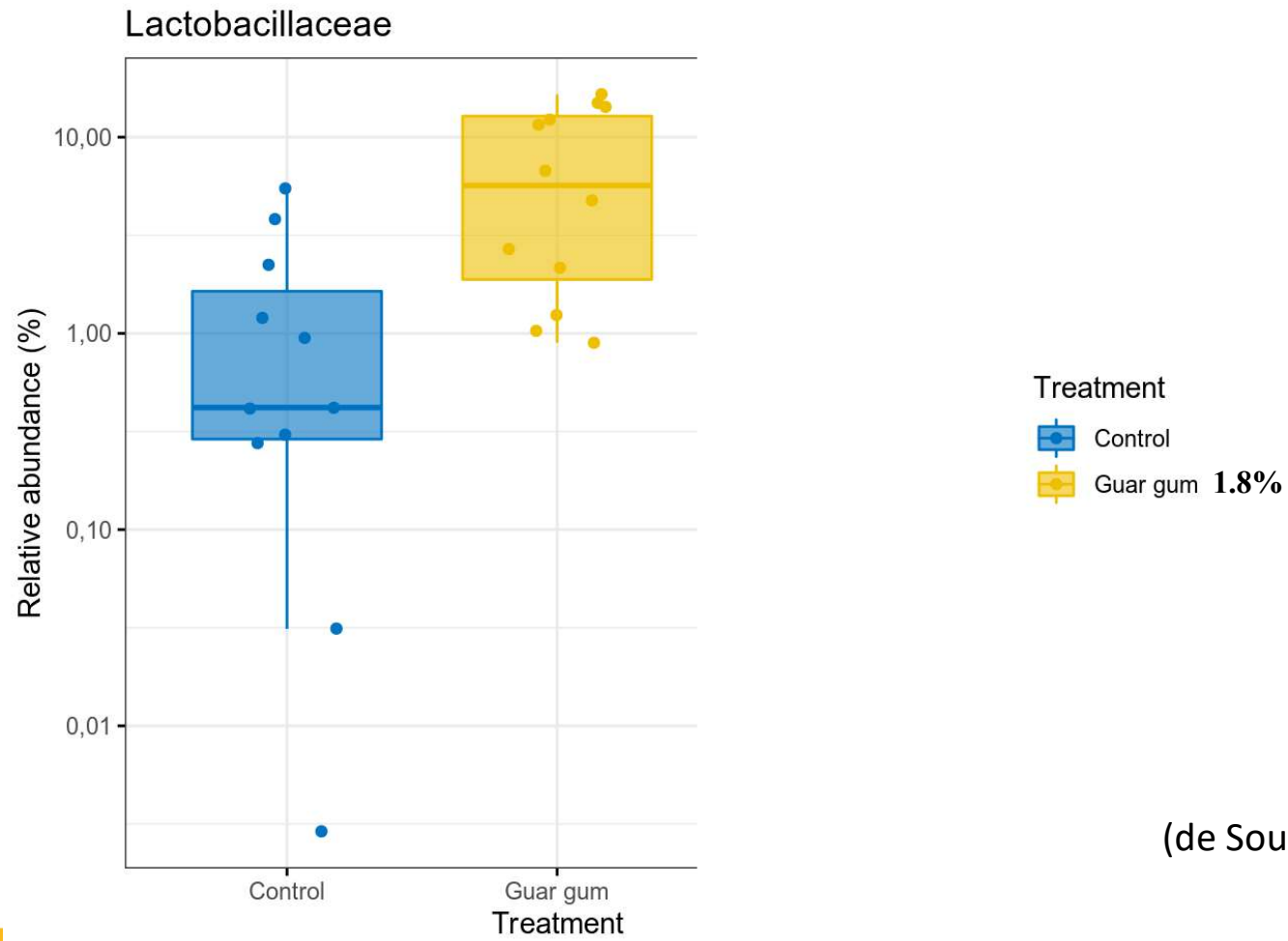
Mannan oligosaccharides (MOS)

## Caecum day 28



(de Souza et al., in press)

## Caecum day 28



(de Souza et al., in press)



Article

# Metabolic Conversions by Lactic Acid Bacteria during Plant Protein Fermentations


Wim Engels <sup>1,\*</sup>, Jamie Siu <sup>1,2</sup>, Saskia van Schalkwijk <sup>1</sup>, Wilma Wesselink <sup>1</sup>, Simon Jacobs <sup>1</sup>  
and Herwig Bachmann <sup>1,3</sup>

*Foods* **2022**, *11*, 1005. <https://doi.org/10.3390>.

## RESEARCH PAPER



## Acids produced by lactobacilli inhibit the growth of commensal *Lachnospiraceae* and S24-7 bacteria

Emma J. E. Brownlie\*, Danica Chaharlangi\*, Erin Oi-Yan Wong, Deanna Kim, and William Wiley Navarre 

Department of Molecular Genetics, University of Toronto, Toronto, ON, Canada

### ABSTRACT

The *Lactobacillaceae* are an intensively studied family of bacteria widely used in fermented food and probiotics, and many are native to the gut and vaginal microbiota of humans and other animals. Various studies have shown that specific *Lactobacillaceae* species produce metabolites that can inhibit the colonization of fungal and bacterial pathogens, but less is known about how *Lactobacillaceae* affect individual bacterial species in the endogenous animal microbiota. Here, we show that numerous *Lactobacillaceae* species inhibit the growth of the *Lachnospiraceae* family and the S24-7 group, two dominant clades of bacteria within the gut. We demonstrate that inhibitory activity is a property common to homofermentative *Lactobacillaceae* species, but not to species that use heterofermentative metabolism. We observe that homofermentative *Lactobacillaceae* species robustly acidify their environment, and that acidification alone is sufficient to inhibit growth of *Lachnospiraceae* and S24-7 growth, but not related species from the *Clostridiales* or *Bacteroidales* orders. This study represents one of the first in-depth explorations of the dynamic between *Lactobacillaceae* species and commensal intestinal bacteria, and contributes valuable insight toward deconvoluting their interactions within the gut microbial ecosystem.

### ARTICLE HISTORY

Received 28 September 2021

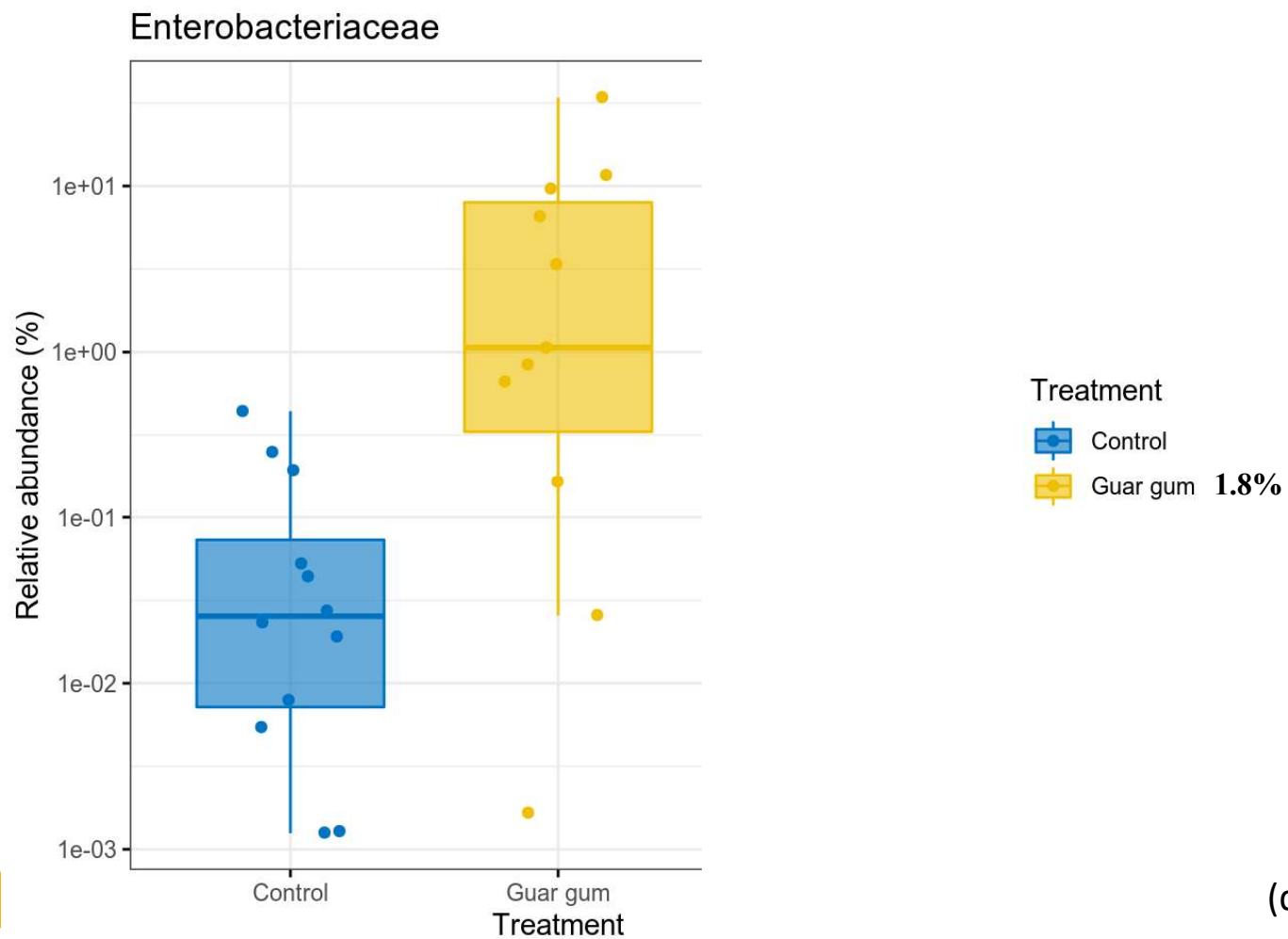
Revised 9 February 2022

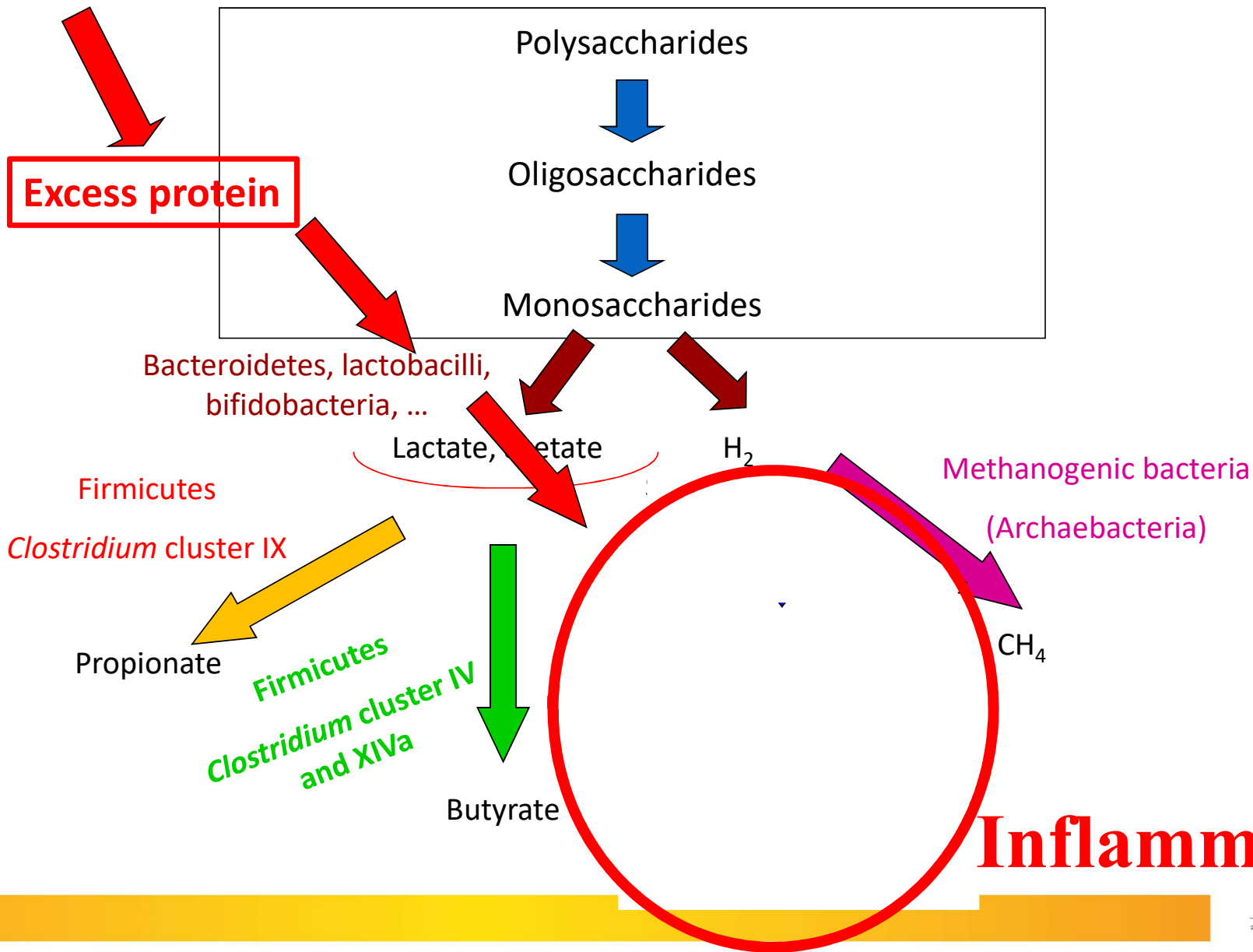
Accepted 18 February 2022

### KEYWORDS

Probiotics; lactobacilli; lactic acid bacteria; *Bacteroidales*; *Clostridiales*; microbiota; gut; acid stress; *Lachnospiraceae*; *Muribaculaceae*; S24-7

Ileum day 14





# Take home messages

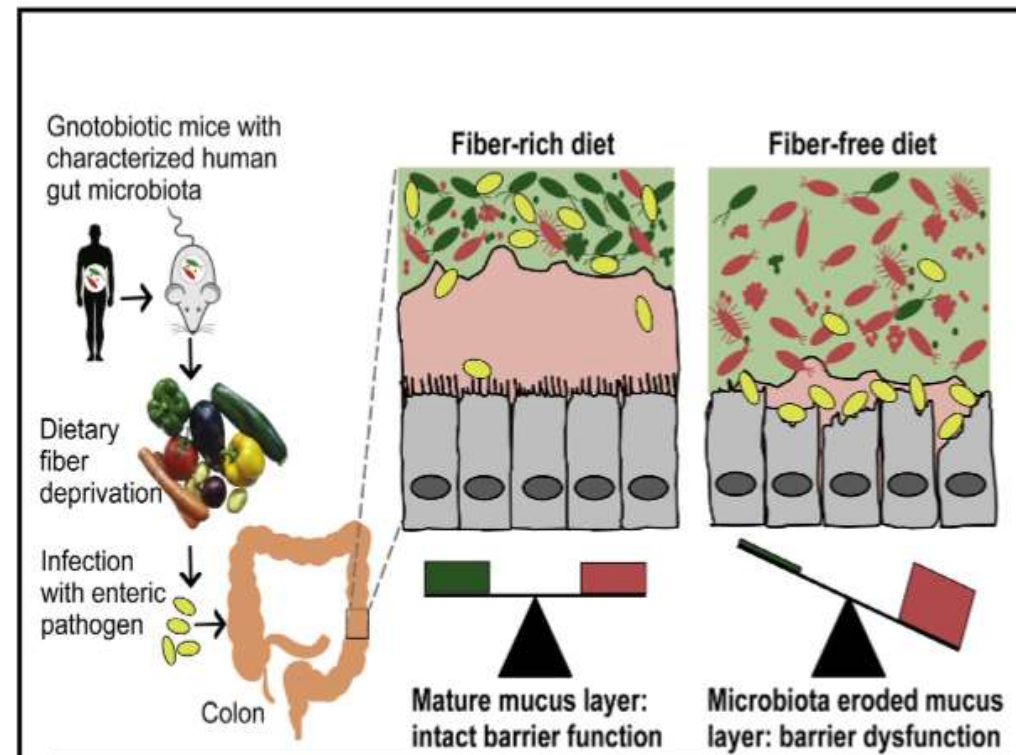
- \* Microbial activity in the caeca determines the efficacy of digestion and absorption in the small intestine
- \* Most broiler feed formulas lack the necessary NSP to support the caecal microbiota
- \* Microbiota of young broilers have difficulty to degrade the complex network of plant cell wall polysaccharides
- \* Added pure NSP, prebiotics, enzymes, etc all can support the caecal microbial network
- \* Proteolytic expansion of Proteobacteria should be avoided

and....



# A Dietary Fiber-Deprived Gut Microbiota Degrades the Colonic Mucus Barrier and Enhances Pathogen Susceptibility

## Graphical Abstract



## Authors

Mahesh S. Desai, Anna M. Seekatz,  
Nicole M. Koropatkin, ...,  
Thaddeus S. Stappenbeck,  
Gabriel Núñez, Eric C. Martens

## Correspondence

mahesh.desai@lih.lu (M.S.D.),  
emartens@umich.edu (E.C.M.)

## In Brief

Regular consumption of dietary fiber helps prevent erosion of the intestinal mucus barrier by the gut microbiome, blunting pathogen infection and reducing the incidence of colitis.

# Thank you for your attention

LIVESTOCK GUT HEALTH TEAM GHENT

[richard.ducatelle@ugent.be](mailto:richard.ducatelle@ugent.be)



## VETERINARY SCIENCES



UNIVERSITEIT  
GENT



FACULTY OF  
VETERINARY MEDICINE  
accredited by EAEVE

